GeneAmp® PCR System 9700

Base Module

P/N N805-0200

User's Manual

Applied Biosystems

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Base Module

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1 Introduction and Safety

Overview

About This Chapter

This chapter provides information to help you safely operate the GeneAmp® PCR System 9700.

In This Chapter

The following topics are covered in this chapter:

Topics	See Page
About This Manual	1-2
Instrument Safety	1-3
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GeneAmp PCR System 9700 Labels and Warnings	1-9
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About This Manual

Overview

This manual describes how to use the GeneAmp® PCR System 9700. It includes the following chapters and appendixes:

- Chapter 1, “Introduction and Safety,” contains safety information.
- Chapter 2, “Product Overview,” describes the instrument, its components, and requirements for installation.
- Chapter 3, “Instrument Setup,” provides information on how to set up the instrument and place it correctly in the laboratory, how to configure the instrument, and how to connect and configure a printer.
- Chapter 4, “Running PCR Samples,” provides information on selecting a method, starting and stopping a run, reviewing the history of a run, and what to do when the run is completed.
- Chapter 5, “Creating and Editing Methods,” describes how to create and edit PCR methods and how to work with stored methods.
- Chapter 6, “Converting Hold Times,” provides information about setting hold times for the GeneAmp® PCR System 9700 compared to the DNA Thermal Cycler or DNA Thermal Cycler 480.
- Chapter 7, “Routine Maintenance,” provides procedures for routine maintenance on the instrument.
- Chapter 8, “Troubleshooting,” lists error messages and suggestions for dealing with other problems you may encounter.
- Appendix A, “Instrument Specifications,” describes the dimensions, power, and electrical specifications of the GeneAmp® PCR System 9700 system, including the control panel, sample temperature information, and printer specifications.
- Appendix B, “Supplied Methods,” contains information about the methods that are supplied with the instrument.
- Appendix C, “Contacting Services and Support,” explains how to contact Applied Biosystems’ Technical Support staff.
- Appendix D, “Limited Warranty Statement,” contains the instrument warranty statement.

Instrument Safety

Before Operating the Instrument

Ensure that everyone involved with the operation of the instrument has:

- Received instruction in general safety practices for laboratories
- Received instruction in specific safety practices for the instrument
- Read and understood all related MSDSs

CAUTION Avoid using this instrument in a manner not specified by Applied Biosystems. Although the instrument has been designed to protect the user, this protection can be impaired if the instrument is used improperly.

Documentation User Attention Words

Five user attention words appear in the text of all Applied Biosystems user documentation. Each word implies a particular level of observation or action as described below.

Note Calls attention to useful information.

IMPORTANT Indicates information that is necessary for proper instrument operation.

CAUTION Indicates a potentially hazardous situation which, if not avoided, may result in minor or moderate injury. It may also be used to alert against unsafe practices.

WARNING Indicates a potentially hazardous situation which, if not avoided, could result in death or serious injury.

DANGER Indicates an imminently hazardous situation which, if not avoided, will result in death or serious injury. This signal word is to be limited to the most extreme situations.

Symbols on Instruments

Electrical Symbols

The following chart is an illustrated glossary of all electrical symbols that are used on Applied Biosystems instruments. Whenever such symbols appear on instruments, please observe appropriate safety procedures.

	Indicates the **On** position of the main power switch.
	Indicates the **Off** position of the main power switch.
	Indicates the **On/Off** position of a push-push main power switch.
	Indicates a terminal that may be connected to the signal ground reference of another instrument. This is not a protected ground terminal.
	Indicates a protective grounding terminal that must be connected to earth ground before any other electrical connections are made to the instrument.
	Indicates a terminal that can receive or supply alternating current or voltage.
	Indicates a terminal that can receive or supply an alternating or direct current or voltage.

Safety Symbols

The following is an illustrated glossary of all nonelectrical safety alert symbols found on Applied Biosystems instruments. Each symbol may appear by itself or in combination with text that explains the relevant hazard (see “Safety Labels on Instruments” on page 1-5). These safety symbols may also appear next to DANGERS, WARNINGS, and CAUTIONS that occur in the text of this and other product-support documents.

Symbol	Description
	Indicates that you should consult the manual for further information and to proceed with appropriate caution.
	Indicates the presence of an electrical shock hazard and to proceed with appropriate caution.
	Indicates the presence of a hot surface or other high-temperature hazard and to proceed with appropriate caution.

Symbol	Description
	Indicates the presence of a laser inside the instrument and to proceed with appropriate caution.
	Indicates the presence of moving parts and to proceed with appropriate caution.

Safety Labels on Instruments

The following CAUTION, WARNING, and DANGER statements may be displayed on Applied Biosystems instruments in combination with the safety symbols described in the preceding section.

English	Francais
CAUTION Hazardous chemicals. Read the Material Safety Data Sheets (MSDSs) before handling.	**ATTENTION** Produits chimiques dangeureux. Lire les fiches techniques de sûreté de matériels avant la manipulation des produits.
CAUTION Hazardous waste. Read the waste profile (if any) in the site preparation guide for this instrument before handling or disposal.	**ATTENTION** Déchets dangereux. Lire les renseignements sur les déchets avant de les manipuler ou de les éliminer.
CAUTION Hazardous waste. Refer to MSDS(s) and local regulations for handling and disposal.	**ATTENTION** Déchets dangereux. Lire les fiches techniques de sûreté de matériels et la régulation locale associées à la manipulation et l'élimination des déchets.
WARNING Hot lamp.	**AVERTISSEMENT** Lampe brûlante.
WARNING Hot. Replace lamp with an Applied Biosystems lamp.	**AVERTISSEMENT** Composants brûlants. Remplacer la lampe par une lampe Applied Biosystems.
CAUTION Hot surface.	**ATTENTION** Surface brûlante.
DANGER High voltage.	**DANGER** Haute tension.
WARNING To reduce the chance of electrical shock, do not remove covers that require tool access. No user-serviceable parts are inside. Refer servicing to Applied Biosystems qualified service personnel.	**AVERTISSEMENT** Pour éviter les risques d'électrocution, ne pas retirer les capots dont l'ouverture nécessite l'utilisation d'outils. L'instrument ne contient aucune pièce réparable par l'utilisateur. Toute intervention doit être effectuée par le personnel de service qualifié de Applied Biosystems.
CAUTION Moving parts.	**ATTENTION** Parties mobiles.

Chemical Safety

Chemical Hazard Warning

WARNING **CHEMICAL HAZARD**. Some of the chemicals used with Applied Biosystems instruments and protocols are potentially hazardous and can cause injury, illness, or death.

- Read and understand the material safety data sheets (MSDSs) provided by the chemical manufacturer before you store, handle, or work with any chemicals or hazardous materials.
- Minimize contact with chemicals. Wear appropriate personal protective equipment when handling chemicals (*e.g.,* safety glasses, gloves, or protective clothing). For additional safety guidelines, consult the MSDS.
- Minimize the inhalation of chemicals. Do not leave chemical containers open. Use only with adequate ventilation (*e.g.*, fume hood). For additional safety guidelines, consult the MSDS.
- Check regularly for chemical leaks or spills. If a leak or spill occurs, follow the manufacturer's cleanup procedures as recommended on the MSDS.
- Comply with all local, state/provincial, or national laws and regulations related to chemical storage, handling, and disposal.

Chemical Waste Hazard Warning

WARNING **CHEMICAL WASTE HAZARD.** Wastes produced by Applied Biosystems instruments are potentially hazardous and can cause injury, illness, or death.

- Read and understand the material safety data sheets (MSDSs) provided by the manufacturers of the chemicals in the waste container before you store, handle, or dispose of chemical waste.
- Handle chemical wastes in a fume hood.
- Minimize contact with chemicals. Wear appropriate personal protective equipment when handling chemicals (*e.g.,* safety glasses, gloves, or protective clothing). For additional safety guidelines, consult the MSDS.
- Minimize the inhalation of chemicals. Do not leave chemical containers open. Use only with adequate ventilation (*e.g.*, fume hood). For additional safety guidelines, consult the MSDS.
- After emptying the waste container, seal it with the cap provided.
- Dispose of the contents of the waste tray and waste bottle in accordance with good laboratory practices and local, state/provincial, or national environmental and health regulations.

About MSDSs

Some of the chemicals used with this instrument may be listed as hazardous by their manufacturer. When hazards exist, warnings are prominently displayed on the labels of all chemicals.

Chemical manufacturers supply a current MSDS before or with shipments of hazardous chemicals to new customers and with the first shipment of a hazardous chemical after an MSDS update. MSDSs provide you with the safety information you need to store, handle, transport and dispose of the chemicals safely.

We strongly recommend that you replace the appropriate MSDS in your files each time you receive a new MSDS packaged with a hazardous chemical.

CHEMICAL HAZARD. Be sure to familiarize yourself with the MSDSs before using reagents or solvents.

Ordering MSDSs

You can obtain from Applied Biosystems the MSDS for any chemical supplied by Applied Biosystems. This service is free and available 24 hours a day.

To obtain MSDSs:

1. Go to **https://docs.appliedbiosystems.com/msdssearch.html**
2. In the Search field, type in the chemical name, part number, or other information that appears in the MSDS of interest. Select the language of your choice, then click **Search**.
3. Find the document of interest, right-click the document title, then select any of the following:
   - **Open** – To view the document
   - **Print Target** – To print the document
   - **Save Target As** – To download a PDF version of the document to a destination that you choose
4. To have a copy of a document sent by fax or e-mail, select **Fax** or **Email** to the left of the document title in the Search Results page, then click **RETRIEVE DOCUMENTS** at the end of the document list.
5. After you enter the required information, click **View/Deliver Selected Documents Now**.

For chemicals not manufactured or distributed by Applied Biosystems, call the chemical manufacturer.

About Waste Disposal

As the generator of potentially hazardous waste, it is your responsibility to perform the actions listed below.

- Characterize (by analysis if necessary) the waste generated by the particular applications, reagents, and substrates used in your laboratory.
- Ensure the health and safety of all personnel in your laboratory.
- Ensure that the instrument waste is stored, transferred, transported, and disposed of according to all local, state/provincial, or national regulations.

Note Radioactive or biohazardous materials may require special handling, and disposal limitations may apply.

GeneAmp PCR System 9700 Labels and Warnings

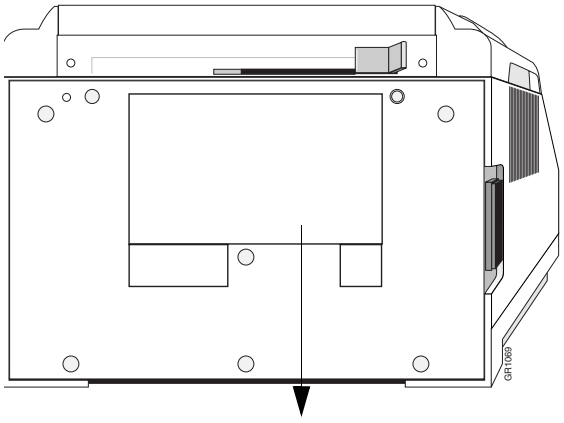
Instrument Safety Labels

Safety labels are located on the instrument. Each safety label has three parts:

- A signal word panel, which implies a particular level of observation or action (*e.g.,* CAUTION or WARNING). If a safety label encompasses multiple hazards, the signal word corresponding to the greatest hazard is used.
- A message panel, which explains the hazard and any user action required.
- A safety alert symbol, which indicates a potential personal safety hazard.

Instrument Warnings Diagram

The following diagram shows the hazards and warnings labels located on the back of the GeneAmp® PCR System 9700 instrument.



Applied Biosystems

GeneAmp™ PCR System 9700

Part No: N8050200

Serial No: 850SYMMDD00

MFG: XXXXXXXXXX XXXX

LISTED

Laboratory Use Electrical Equipment

3277 ; EN61326, Group 1; Class B

C UL US CE N2284

Made in Singapore

Electrical Rating:

100/120/220/230/240 VAC~

50/60 Hz; Max. Power 725 VA

Fuse Rating:

250 VAC, 8A, SB(T)

Replace only with same

TYPE and RATING FUSE

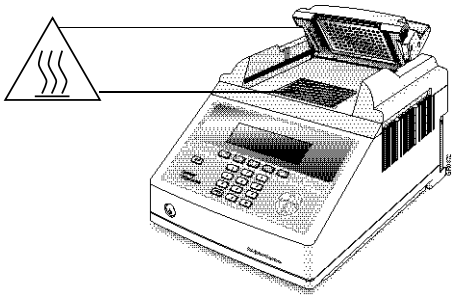
DO NOT OPEN

PANELS OR COVERS

KEEP FREE OF DEBRIS FAN INTAKE

Danger of Burns

CAUTION **PHYSICAL INJURY HAZARD**. Hot Surface. Use care when working around this area to avoid being burned by hot components.



Grounding and Electrical Safety

The system 9700 must be grounded for protection against electrical shock.

DANGER **ELECTRICAL HAZARD.** Do not use an adapter to a two-terminal outlet since this does not provide positive ground protection.

Fuses

Improper fuses can damage the wiring system and cause a fire.

WARNING **FIRE HAZARD.** For continued protection against the risk of fire, replace fuses only with fuses of the type and rating specified for the instrument.

DANGER **ELECTRICAL SHOCK HAZARD.** Improper fuses or high voltage supply can damage the instrument wiring system and cause a fire. Before turning on the instrument, verify that the fuses are properly installed and that the instrument voltage matches the power supply in your laboratory.

Laboratory Environmental Requirements

Introduction

Take the precautions described in this section whenever you operate the system 9700. Read this section before you install the instrument.

CAUTION The instrument should be used according to the instructions provided in this manual. If used otherwise, the protection provided by this instrument may be impaired.

Temperature, Humidity, and Environment

IMPORTANT This instrument is designed for indoor use.

IMPORTANT Do not operate in a Cold Room or a refrigerated area. The system 9700 will operate safely when the ambient temperature is 5 °C to 40 °C (41 °F to 104 °F) and will meet performance specifications when the ambient temperature is 15 °C to 30 °C and the ambient relative humidity is 20 to 80%. These specifications have been calculated for altitudes between 0 and 2,000 meters.

CAUTION **FIRE HAZARD.** This instrument is not designed for operation in an explosive environment. Do not place the instrument close to potentially explosive materials or objects.

IMPORTANT The instrument should be stored between –20 °C and 60 °C (–4 °F and 140 °F) at altitudes between 0 and 12,000 meters.

Note This instrument is able to withstand transient overvoltage according to Installation Category II as defined in IEC 1010-1.

Pollution

The installation category (overvoltage category) for this instrument is II, and it is classified as portable equipment. The instrument has a pollution degree rating of 2 and may be installed in an environment that has non-conductive pollutants only.

Emission/Immunity Statement

For our European customers, any product marked with the CE label meets the European EMC directive 89/336/EEC and the Low Voltage Directive 72/23/EEC. This product meets Class B emission limits.

Routine Maintenance for Safe Operation

If you use any cleaning or decontamination method, except those recommended in the manual, you may risk damaging the equipment.

Maintain your instrument in good working order. In the event that the instrument has been subjected to adverse environmental conditions (such as fire, flood, earthquake, etc.), contact your local sales office for advice.

Safety and Electromagnetic Compatibility (EMC) Standards

This section provides information on:

- U.S. and Canadian Safety Standards
- Canadian EMC Standard
- European Safety and EMC Standards
- Australian EMC Standards

U.S. and Canadian Safety Standards

This instrument has been tested to and complies with standard UL 3101-1, "Safety Requirements for Electrical Equipment for Laboratory Use, Part 1: General Requirements."

This instrument has been tested to and complies with standard CSA 1010.1, "Safety Requirements for Electrical Equipment for Measurement, Control, and Laboratory Use, Part 1: General Requirements."

Canadian EMC Standard

This instrument has been tested to and complies with ICES-001, Issue 3: Industrial, Scientific, and Medical Radio Frequency Generators.

European Safety and EMC Standards

Safety

This instrument meets European requirements for safety (Low Voltage Directive 73/23/EEC). This instrument has been tested to and complies with standards EN 61010-1:2001, "Safety Requirements for Electrical Equipment for Measurement, Control and Laboratory Use, Part 1: General Requirements" and EN 61010-2-010, "Particular Requirements for Laboratory Equipment for the Heating of Materials."

EMC

This instrument meets European requirements for emission and immunity (EMC Directive 89/336/EEC). This instrument has been tested to and complies with standard EN 61326 (Group 1, Class B), "Electrical Equipment for Measurement, Control and Laboratory Use – EMC Requirements."

Australian EMC Standards

This instrument has been tested to and complies with standard AS/NZS 2064, "Limits and Methods Measurement of Electromagnetic Disturbance Characteristics of Industrial, Scientific, and Medical (ISM) Radio-frequency Equipment."

Product Overview 2

Overview

About This Chapter This chapter describes the GeneAmp® PCR System 9700, its components, and requirements for installation.

In This Chapter The following topics are contained in this chapter:

Topic	See page
About the GeneAmp PCR System 9700	2-2

About the GeneAmp PCR System 9700

About the Instrument

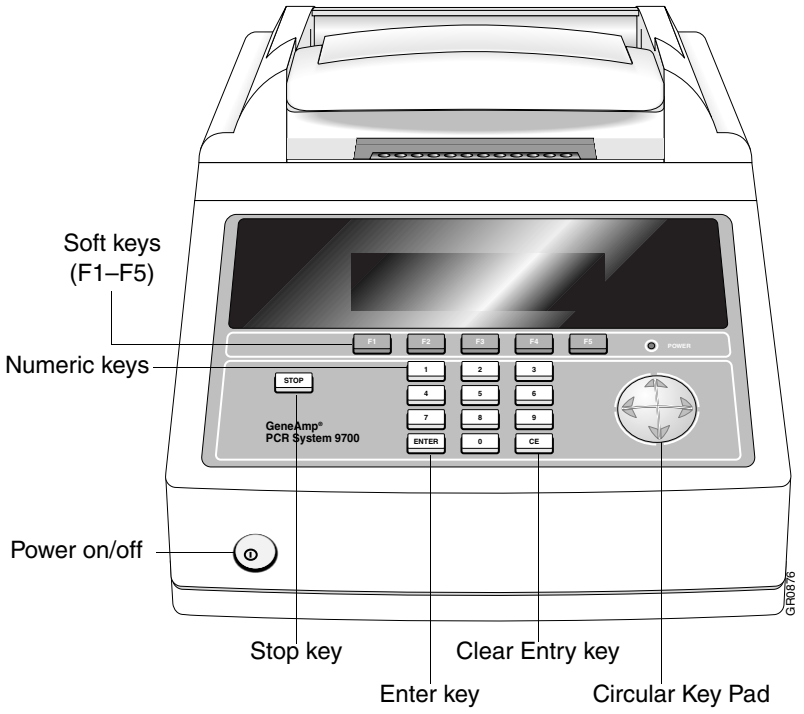
The GeneAmp PCR System 9700 is an automated instrument, specifically designed for the amplification of nucleic acids using the Polymerase Chain Reaction (PCR) process. The user interface consists of a control panel with a full numeric keypad, soft keys, and a graphical display screen that shows the time and temperature profile for each run.

Control Panel

The instrument control panel consists of a display screen and 22 keys. The display screen shows a graphical representation of PCR events, including pre-PCR holds, PCR cycling, and post-PCR holds. You use the keys to enter information into fields on the display screen.

Control Panel Diagram

The following diagram shows the control panel.



Using the Keys

The following table describes the Control Panel keys in the previous figure.

Key	Use to...
Soft keys (F1-F5)	Select the function specified above the key. The function of each key is defined on the display screen above the key, and is redefined as you view different screens.
Numeric keys	Enter numbers from left to right into a field you highlight.
Stop key	Stop a method while it is running.
Enter key	Enter information typed into a field and advance the highlight box to the next field on a screen.
Clear Entry key (CE)	Remove information from a field.
Circular Key Pad	Move the highlight box to different fields on the display screen in the direction of the arrow.

Selecting a Field

There are two ways to select a field.

If you want to...	Then....
move the highlight box in one of four directions	Use the Circular Key Pad.
advance the box to the next field	Press the Enter key.

Entering Numeric Values

The following table lists how to enter numeric values for the Temperature and Hold Time parameters.

Parameter	Description
Temperature parameters	Enter values for temperature in decimal form. It is not necessary to type a decimal point. For example, to specify 89.0 °C, press 8 9 0, then press Enter.
Hold Time parameters	A hold time is the length of time the samples will be maintained at a specified temperature. Specify all hold times in minutes and seconds, then press Enter. For example, to specify one minute and five seconds, press 1 0 5, then press Enter.

Transfer or Storage of Methods

You can transfer or store methods using a Methods Storage Card. The instrument can run a method from its own software or from the methods stored on a Methods Storage card.

Use...	To...	See...
Methods Storage Card, Centennial 256kb SRAM (P/N 0940-1064)	transfer methods from the Methods Storage card to the instrument or from the instrument to the card for storage.	“Copying Methods” on page 3-24.

Upgrading the Firmware

There are two ways to upgrade the firmware:

- With a serial cable connection (PC communication cable P/N N805-1327 or Macintosh® communication cable P/N N805-1328) from the instrument’s RS485 ports
- With a PCMCIA Flash Memory Card

Note Upgrade firmware can be ordered from the Applied Biosystems web site or by contacting PCR Technical Support.

For more information about upgrading the firmware, see the following.

If you are using...	See...
a serial cable connection (RS485 ports)	“Upgrading Through the Serial Port” on page 3-18.
a PCMCIA Flash Memory Card	“Upgrading Firmware from a PCMCIA Flash Memory Card” on page 3-22.
Microsoft Windows® 95 or Windows NT®	“Downloading Firmware Using Windows 95 or Windows NT” on page 3-20.
Windows® 3.1	“Downloading Firmware Using Windows 3.1” on page 3-21.

How the 9700 Differs from the GeneAmp PCR System 9600

The GeneAmp PCR System 9700 differs from the GeneAmp® PCR System 9600 in the following ways:

- Reaction volumes of up to 50 µL only can be run and in the “9600 Mode”, as opposed to 100 µL in the GeneAmp PCR System 9600.
- Methods are stored under a user name, allowing users to keep track of and protect their own methods.
- Hold, Cycle, and Auto programs no longer exist as menu items under the Create function. Instead of linking programs together to create a method, you create a single method on the GeneAmp PCR System 9700.
- If the allowed pause time elapses during a manual pause of a method, the method will continue running (instead of stopping).
- The last method run cannot be re-run or edited unless it is first saved.
- The history file no longer includes individual setpoint time and temperature information.
- An automated restart or incubate function is available for use after a power outage.

3 Instrument Setup

Overview

About This Chapter

This chapter provides information on how to set up the GeneAmp® PCR System 9700 and place it correctly in the laboratory, how to configure the instrument, and how to connect and configure a printer.

In This Chapter

The following topics are covered in this chapter:

Topic	See page
Unpacking the Instrument	3-2
Setting Up your Laboratory	3-3
Double-Line Fuse	3-5
Turning On the Instrument	3-6
Setting Custom Parameters	3-7
Running Instrument Diagnostics	3-13
Using the T_m Calculator	3-17
Upgrading the System Firmware	3-18
Copying Methods	3-24
Connecting and Configuring a Printer	3-26

Unpacking the Instrument

Inspecting the Instrument

Inspect the GeneAmp® PCR System 9700, and all other supplied parts, for any damage that may have occurred during shipment. If there has been any damage during transit, notify the carrier and Applied Biosystems immediately.

Note Save the shipping container and all packing materials in case it becomes necessary to reship the instrument.

To order additional instruments or supplies, contact one of the regional offices listed in Appendix C, “Contacting Services and Support.”

Additional Part Numbers

You can order modules, accessories, and disposables from Applied Biosystems.

To order this part...	Use P/N...
GeneAmp PCR System 9700 Base Module	N805-0200
60-Well 0.5 mL Sample Block Module	4309131
Auto-Lid Sample Block Module	4312904
96-Well Gold Sample Block Module	4314443
96-Well Aluminum Sample Block Module	4314445
Dual 384-Well Sample Block Module	N805-0400
Temperature Verification System	N801-0435
PC (method storage) card	940-1064
Printer cable	N805-1326
PC communication cable	N805-1327
Macintosh® communication cable	N805-1328

Note See the *Interchangeable Sample Block Module Users Manual* for part numbers of disposable items.

Setting Up your Laboratory

Choosing a Location

To prevent vibration, place the GeneAmp PCR System 9700 on a solid, stable, level surface that allows free airflow overhead and around the sides and back. You should keep all ventilation slots in the instrument cover free of obstruction, for example from excess printer paper.

Operating Temperature

The instrument will meet performance specifications when the ambient temperature is 15 °C to 30 °C (59 ° to 86 °Fahrenheit) and the ambient relative humidity is 20% to 80%.

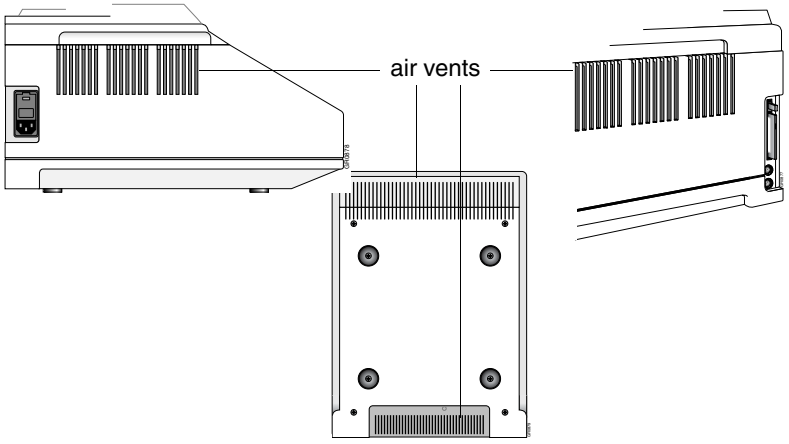
What to Avoid

When setting up the instrument, avoid:

- Placing the instrument under overhanging shelves, especially when there is a wall behind the unit.
- Proximity to other instruments on the same bench or other heat-generating equipment.
- Locations subject to wide temperature fluctuation, such as direct sunlight, or air drafts.
- Damp areas.
- Environments where there is an oil mist.

Do Not Block Air Vents

Do not block the circulation of air to the vents located on the sides and bottom of the instrument



Note When operating multiple 9700 instruments, provide at least 8 in. of space in-between the instruments.

Fuse Service Configuration

The instrument is shipped with double-line service configuration.

DANGER **ELECTRICAL SHOCK HAZARD**. To protect yourself against shock hazards, use a properly wired three-terminal outlet. Do not use an adapter to a two-terminal outlet.

WARNING **FIRE HAZARD**. Improper fuses can damage the wiring system and cause a fire. Before turning on the instrument, verify that the fuses are properly installed.

Input Voltage

IMPORTANT You must be able to disconnect the main power supply to the instrument immediately if necessary.

The following table specifies the electrical operating range for the instrument in various parts of the world. Select the appropriate fuse configuration based on the voltage used.

Location	Voltage (VAC)[a]	Frequency	Amperage (A) Nominal
Japan	100 ±10%	50/60 Hz ±1%	3.16
USA/Canada	120 ±10%	50/60 Hz ±1%	4.20
EC	230 ±10%	50/60 Hz ±1%	3.14

a. Acceptable AC line voltage tolerances: 100, 120, 220, 230 ±10%; 240 VAC +6%/–10%, 50/60 Hz ± 1%.

Note The Volt-Amp number for this instrument is 725 Volt Amps.

Interchangeable Sample Block Modules

The instrument features an interchangeable sample block module allowing portability and versatility in sample configuration.

Sample Block Module	Part Number
96-Well Gold Sample Block Module	4314443
96-Well Aluminum Sample Block Module	4314445
60-Well 0.5 mL Sample Block Module	4309131
Dual 384-Well Sample Block Module	N805-0400
Auto-Lid Sample Block Module	4312904

Note The GeneAmp PCR System 9700 will not operate without an interchangeable sample block module installed.

For an example, refer to the *GeneAmp PCR System 9700 96-Well Sample Block Module User Guide*, for instructions on how to install a block and configure the samples.

Double-Line Fuse

Introduction

WARNING **FIRE HAZARD**. For continued protection against the risk of fire, replace fuses only with listed and certified fuses of the same type and rating as those currently in the instrument.

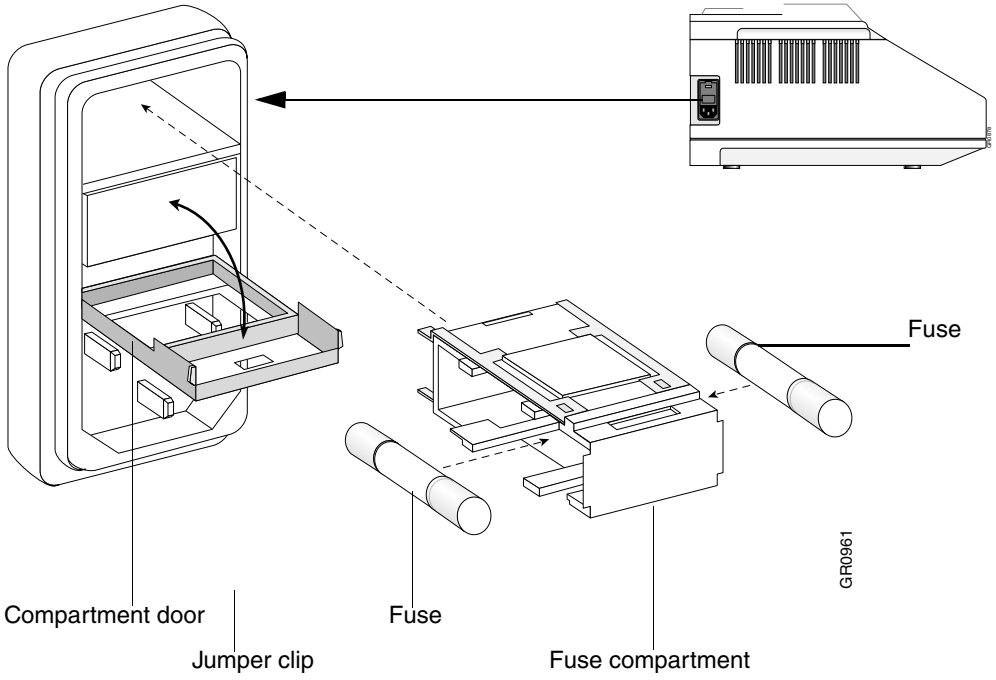
IMPORTANT For proper operation, you must know the power source(s) available and determine if the instrument fuse configuration is correct. The instrument is shipped configured for single-line operation.

Fuse Part Number

The correct fuse is an 8 amp Type T 250 V 5x20 mm fuse, P/N 0999-1683.

Power Entry Module Diagram

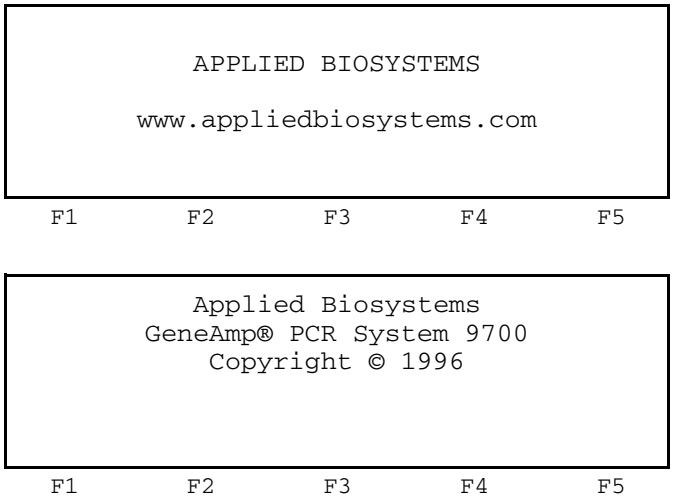
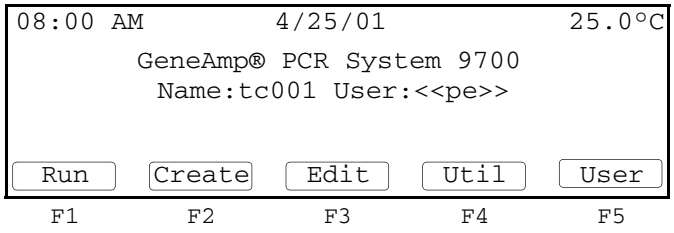
The following is a diagram of the Power Entry Module.



Turning On the Instrument

Turning On the Instrument

To turn on the instrument:

Step	Action
1	Plug the power cord into the side of the instrument and into an outlet.
2	Press the power switch to the **ON** position. The cooling fan powers up, and the Start-up screens appear. 
3	After several seconds the **Main** menu appears. You can use any of the functions displayed above the soft keys.  **Note** The Main menu should appear within a few seconds. If any permanent patterns of lines or bars display on the screen, contact Applied Biosystems Technical Support.

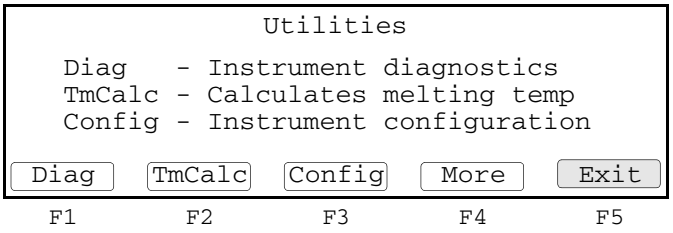
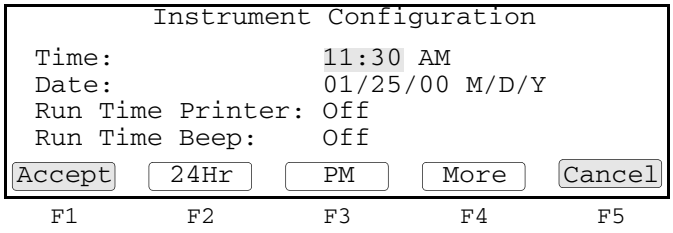
Setting Custom Parameters

Overview

The instrument is shipped with default configuration values. The following procedure describes how to set customized values as well as how to enable or disable optional features.

Displaying the Configuration Screen

To display the Configuration Screen:

Step	Action
1	From the **Main** menu, press **Util**. The Utilities screen appears.  From the Utilities screen, you can take the following action: **If you want to...** — **See page** Delete a method — 5-20 Run instrument diagnostics — 3-13 Review the history of a run — 4-10 Access the T_m calculator — 3-17 Copy methods to or from a methods transportability card — 3-24
2	Press **Config**. The first Configuration screen appears. 

Setting the Time

In the first Configuration screen, you can set the current time and date for file memory maintenance, and run-time displays. You can also enable or disable the run-time printer and the run-time beep.

To set the time:

Step	Action
1	Use the Circular Key Pad to select the **Time** field.
2	Press the **24 Hr** or **PM (AM)** soft keys until the format you want for the current time displays in the Time field.
3	Use the numeric keys to type in the hours followed by minutes.
4	Press **Accept** when your entries are complete. **Note** **CE** clears an entry.

Setting the Date

You must set the instrument to the correct date.

To set the date:

Step	Action
1	Use the circular key to select the **Date** field.
2	The three fields to set in the **Date** field are the: ♦ Days field ♦ Month field ♦ Year field
3	Press the **D/M/Y** or **Y/M/D** soft keys until the format you want for the current date displays in the Date field.
4	Use the numeric keys and type in a number for each of the three fields. The order of these three fields depends on the format you chose in step 3.

Enabling or Disabling a Printer

Enabling the printer allows you to print method parameters or records of run time events directly from the display screen. The default value for the optional printer is Off.

Note For connection and configuration see “Connecting and Configuring a Printer” on page 3-26.

To enable or disable the optional printer:

Step	Action
1	Select the **Run Time Printer** field. This changes the functions of the soft keys.
2	You can take the following action: **If you want to...** / **Then...** enable the printer / Press ON. disable the printer / Press Off.
3	Press **Enter** to accept your entry.
4	Press **Cancel** to cancel all entries and return to the previous screen.

Turning the Run-Time Beeper On or Off

When turned on, the run-time beeper beeps during a pause and once at the completion of a run. The default value is Off.

To turn the Run Time-Beeper on or off:

Step	Action
1	Select the **Run Time Beep** field. This changes the value of the soft keys.
2	You can take the following action: **If you want to...** / **Then...** turn the beeper on / Press ON. turn the beeper off / Press Off.
3	Press **Enter** to accept your entry.

Setting the Pause Time Out

The Pause Time Out field sets the time in minutes:seconds format for the length of a pause when you use the Pause soft key to pause a run from the Run Time screen. For more information on Pausing a Run, see “Pausing a Run” on page 4-8.

To set the pause time out:

Step	Action
1	Select **More** to display the second configuration screen. Instrument Configuration Pause Time Out: 10:00 (00:01-99:59) Idle State Setpoint: 25.0°C (4.0-99.9) Baud Rate: 9600 Accept More Cancel F1 F2 F3 F4 F5
2	Set the **Pause Time Out** field by using the numeric keys to enter a time in minutes and seconds. From this screen, you can also define the: ♦ Idle state setpoint temperature. ♦ Baud rate for your printer port.
3	Press **Accept** when all information on this screen is correct. **Note** **CE** clears an entry.

Defining the Idle State Setpoint Temperature

The Idle State Set Point temperature is the temperature at which the instrument will remain when powered up, but idle.

IMPORTANT After a run is completed or terminated, there is approximately a 30 second delay before the instrument attains the specified idle state temperature. This allows you to stop one method and start another before the instrument temperature changes.

To define the Idle State Set Point temperature:

Step	Action
1	Select the **Idle State Set Point** field.
2	Use the numeric keys and type in a temperature between 4.0° C and 99.9 °C. **Note** The default is 25 °C.
3	Press **Enter** to accept your entry.

Defining the Baud Rate for Your Printer Port

The following section contains a table that lists serial board specifications and a procedure that describes how to define the printer port value in the Baud Rate field.

Serial Board Specifications

You can connect the GeneAmp PCR System 9700 to any printer with a serial interface board and the following specifications.

Baud Rate	9600
Parity	NONE
Data Bits	8
Stop Bits	1

How to Set the Baud Rates

To set baud rates:

Step	Action
1	Select the **Baud Rate** field.
2	You can take the following action: **If you want to...** / **Then...** increase the baud rate / Press Up. decrease the baud rate / Press Down. **Note** Available baud rates are 19200, 9600, 4800, 1200, 600, and 300.
3	Press **Enter** to accept your entry.

Configuring the Contrast and Screen Saver Options

You can use the third Configuration screen to set:

- Screen contrast from 1 to 20 (light to dark)
- Screen saver mode

Displaying the Screen

From the second Configuration screen, choose More to display the third Configuration screen.

Screen Saver Options

The following table lists the screen saver options:

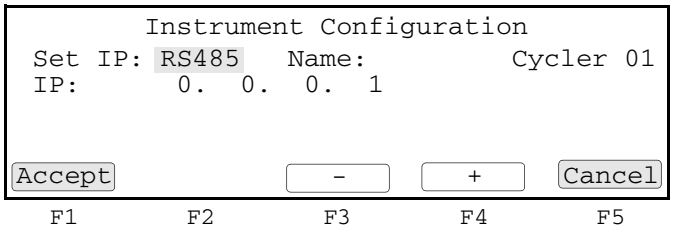
Use This Option...	To Activate the Screen Saver...
Smart	After 15 minutes unless a method is running
Always	After 15 minutes
Never	Never

Setting the IP Address

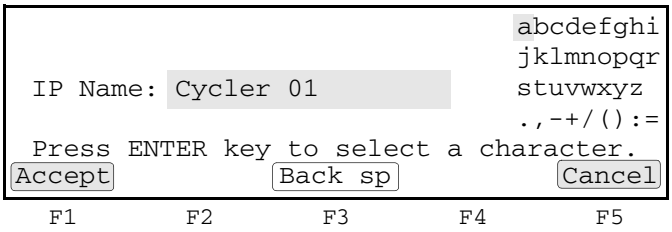
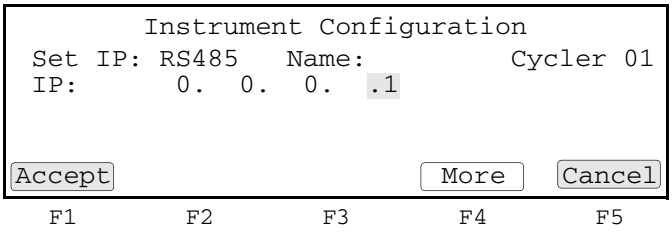
You can use the fourth Configuration screen to set the IP address for the instrument. The instrument can be linked to a network via the RS485 ports located on the right-rear side of the instrument (see the figure on page 3-18). Once connected and configured, the instrument will be visible from the network under the IP address and instrument name you select in the following procedure.

Note See the *System 9700 Networking Software User's Manual* (P/N 4309575) for more information about networking GeneAmp PCR System 9700 instruments.

To set the IP Address for an instrument:

Step	Action
1	Select **More** to display the second configuration screen. 
2	Set the **Set IP** field using the + and – keys. The following settings are available: **Setting** / **Definition** RS485 — Activates the RS 485 ports. Must be used with the networking software Off — Turns off the networking capability and turns on the RS232 ports.
3	Use the Circular Key Pad to select the **Name** field.
4	Select **Name**.

To set the IP Address for an instrument: *(continued)*

Step	Action
5	Using the Circular Key Pad and **Enter** key, enter up to a 16 character name for identifying the instrument on the network. 
6	Press **Accept** when finished entering a name. The software returns to the previous screen and displays the new name in the Name field.
7	Use the Circular Key Pad to select the last **IP** field.
8	Set the IP field by using the numeric keys to enter an IP address for the instrument. 
9	Press **Accept** when all information on this screen is correct.

Running Instrument Diagnostics

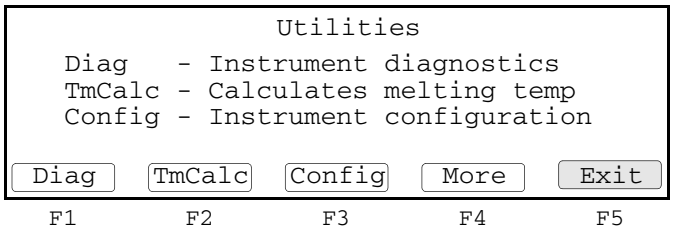
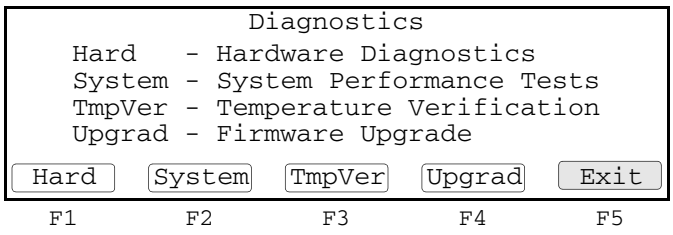
Overview

The instrument provides a number of internal diagnostic utilities, tests you can run to verify that the instrument hardware and software components meet performance specifications.

Viewing the Diagnostic Screen

The following procedure describes how to view the Diagnostic screen.

To view the Diagnostic screen:

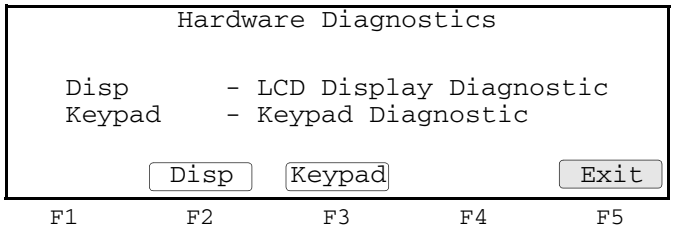
Step	Action
1	From the **Main** menu, press **Util**. The Utilities screen appears. 
2	From the **Utilities** screen, press **Diag**. The Diagnostics screen appears. 
3	Press the soft key that accesses the diagnostic utility you want to run:

Press this soft key...	To...
Hard	Access hardware diagnostic utilities that allow you to test hardware and electrical components such as the Display screen and Keypad.
System	Access the system performance tests. You can run these two system tests to test the rate of system heating and cooling, and the performance of PCR cycling.
TmpVer	Access the temperature verification tests that allow you to verify sample block calibration and uniformity. These two tests are discussed in the users manual for the interchangeable sample block.
Upgrade	Access a utility that allows you to update the 9700 instrument firmware through the RS485 serial port or using a PCMCIA Flash Memory card.

Testing the Electrical Components

Use the hardware diagnostic utilities to test the electrical components of the 9700 instrument.

To test the electrical components:

Step	Action
1	From the **Diagnostics** screen, press **Hard**. The Hardware Diagnostics appears. 
2	Use the corresponding soft key to select the hardware component for which you want to perform one of the following tasks: **Press this soft key...** / **To...** Display / Visually determine if the Liquid Crystal Display (LCD) screen is functioning properly. Keypad / Verify the operation of the keypad.

Testing the Display Screen

The Display diagnostic test allows you to visually determine if the display screen is properly functioning by turning on and off all the LCD pixels.

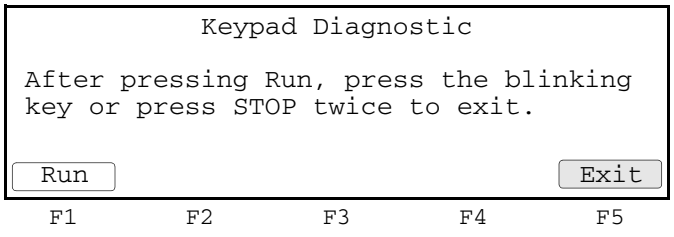
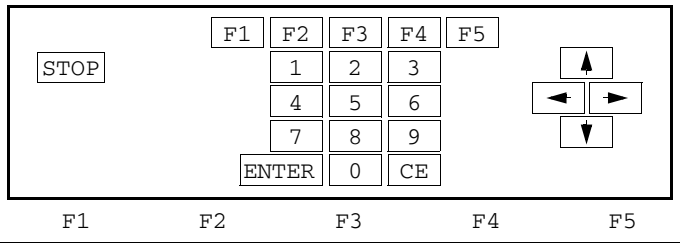
To test the display screen:

Step	Action
1	From the **Hardware Diagnostics** screen, press **Display**. The Display Diagnostic screen appears. Display Diagnostics 1. Read all instructions first. 2. Press Run to turn ON all pixels. 3. Press STOP to turn OFF all pixels. 4. Press STOP to exit. Run Exit F1 F2 F3 F4 F5
2	You can take the following action: **Press...** / **To...** Run / Turn ON all pixels. Check display. Stop / Turn OFF all pixels. Check display. Stop, again / Exit.

Testing the Keypad

Use the Keypad diagnostic test to verify that all 22 keys on the control panel are functioning properly.

To test the keypad:

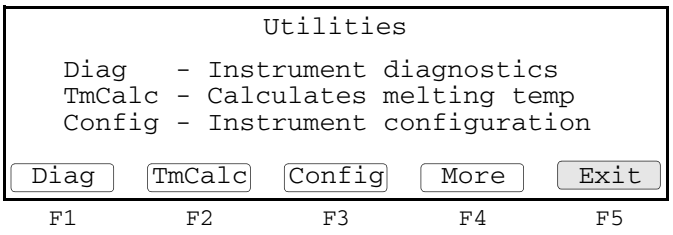
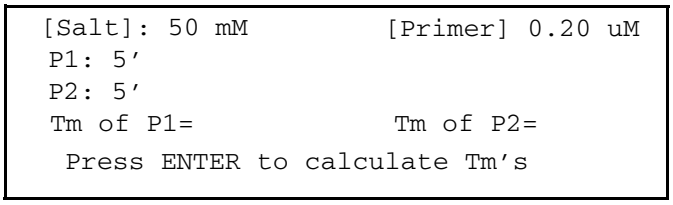
Step	Action
1	From the **Hardware Diagnostics** screen, press **Keypad**. The Keypad Diagnostic screen appears. 
2	Press **Run**. The Control Panel screen appears. 
3	Press the indicated flashing key.

Using the T_m Calculator

How to Use the T_m Calculator

Use the T_m Calculator to determine the annealing temperature of a primer set of known sequence.

To use the T_m Calculator:

Step	Action
1	From the **Main** menu, press **Util**. The Utilities screen opens as shown below. 
2	Press **TmCalc**. The Tm Calculator appears. 
3	Enter the salt concentration. **Note** The default is 50. Enter values 5 to 1000.
4	Enter the primer concentration. **Note** The default it 0.20. Enter values 0.01 to 10.00.
5	Enter primer sequence in P1.
6	Enter primer sequence in P2 and press **Enter** to calculate the T_ms. The melting points are displayed. Use this information to program a run. For more information, see Chapter 5, "Creating and Editing Methods."
7	Press **Return** to display the **Utilities** screen.

Upgrading the System Firmware

Introduction

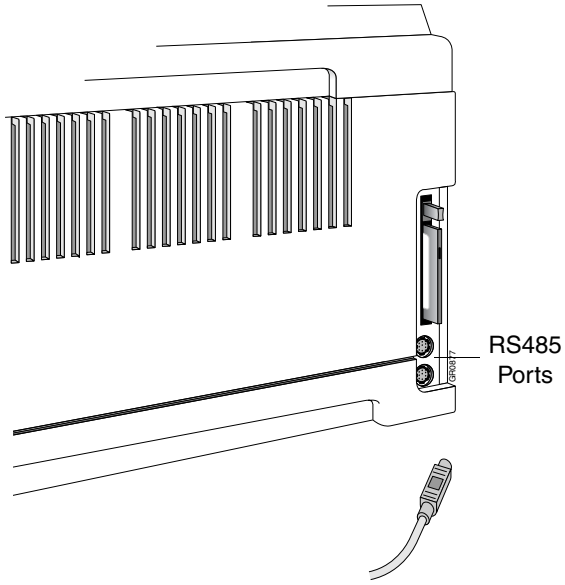
You can update the GeneAmp PCR System 9700 firmware from a serial connection to the RS485 port or from a PCMCIA Flash Memory Card.

Contact your local Applied Biosystems representative to obtain a PCMCIA Flash Memory Card.

Upgrading Through the Serial Port

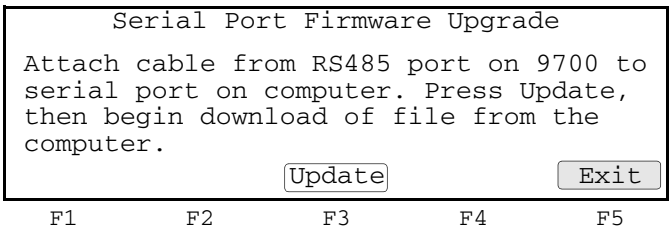
The following procedure describes how to upgrade the firmware through the RS485 serial port.

To upgrade the firmware through the RS485 serial port:

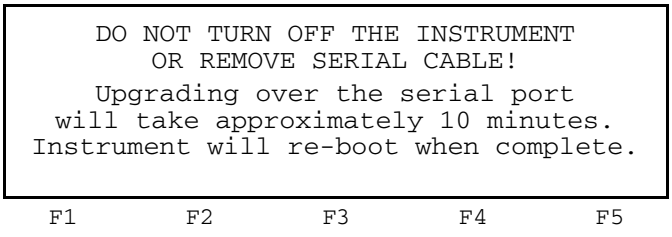
Step	Action
1	Connect the appropriate communication cable from a computer to the RS485 port on the 9700 instrument Base Module (see the figure below). You can use the following cables: **Communication cable** / **Part number** PC communication cable / N805-1327 Macintosh® communication cable / N805-1328 For information on connecting the printer cable, see page 3-26. 
2	From the **Diagnostics** screen, press **Upgrade**. The Firmware Upgrade screen appears. Firmware Upgrade PCcard - Upgrade from PC card Serial - Upgrade from serial port PCcard Serial Exit F1 F2 F3 F4 F5

To upgrade the firmware through the RS485 serial port: *(continued)*

<table>
<tr><th>Step</th><th>Action</th></tr>
<tr><td>3</td><td>From the Firmware Upgrade screen, press Serial.

IMPORTANT Do not remove the cable or turn off the instrument until the upgrade is completed.</td></tr>
<tr><td>4</td><td>From the Serial Port Firmware Upgrade screen, press Update.

 
</td></tr>
<tr><td>5</td><td>Take the following action:

<table>
<tr><th>If you are using...</th><th>See...</th></tr>
<tr><td>Microsoft™ Windows® 95, or Microsoft Windows NT®</td><td>“Downloading Firmware Using Windows 95 or Windows NT” on page 3-20.</td></tr>
<tr><td>Microsoft Windows® 3.1</td><td>“Downloading Firmware Using Windows 3.1” on page 3-21.</td></tr>
</table>
</td></tr>
</table>

Downloading Firmware Using Windows 95 or Windows NT

To download the system firmware using Windows 95 or Windows NT:

<table>
<tr><th>Step</th><th>Action</th></tr>
<tr><td>1</td><td>Click Start > Programs > Accessories > HyperTerminal.</td></tr>
<tr><td>2</td><td>Click the HyperTerminal icon.

The Connection Description dialog box appears.</td></tr>
<tr><td>3</td><td>In the Connection Description dialog box, enter the following information and click OK. The Connect Using dialog box appears.

<table><tr><th>In this field...</th><th>Take this action...</th></tr><tr><td>Name</td><td>Enter a name, for example TEC.</td></tr><tr><td>Icon</td><td>Choose one of the icons.</td></tr></table></td></tr>
<tr><td>4</td><td>In the Connect Using dialog box, connect using either Direct to Com 1 or Direct to Com 2, depending on the port to which the cable is attached.

Note For Windows NT applications this dialog box is titled Connect To, and the fields are COM1and COM2.</td></tr>
<tr><td>5</td><td>Depending on whether you chose Com 1 or Com 2 in the previous step, in the Properties dialog box, enter the following.

<table><tr><th>In this field...</th><th>Enter...</th></tr><tr><td>Bits Per Second</td><td>9600</td></tr><tr><td>Data Bits</td><td>8</td></tr><tr><td>Parity</td><td>None</td></tr><tr><td>Stop Bits</td><td>1</td></tr><tr><td>Flow Control</td><td>None</td></tr></table>
When you have competed entering the information, click OK.</td></tr>
<tr><td>6</td><td>From the Transfer menu, choose Send file.

The Send File dialog box appears.</td></tr>
<tr><td>7</td><td>In the Send File dialog box, enter the following and click Send.

<table><tr><th>In this field...</th><th>Take this action...</th></tr><tr><td>Filename</td><td>Locate the upgrade file with the name under which you saved the file.</td></tr><tr><td>Protocol</td><td>Enter Kermit.</td></tr></table>
A dialog box appears that displays the current packet number and the number remaining. When the download is complete, the GeneAmp PCR System 9700 automatically resets.</td></tr>
<tr><td>8</td><td>From the File menu, choose Save to save the parameters.</td></tr>
</table>

Downloading Firmware Using Windows 3.1

To download the system firmware using Windows 3.1:

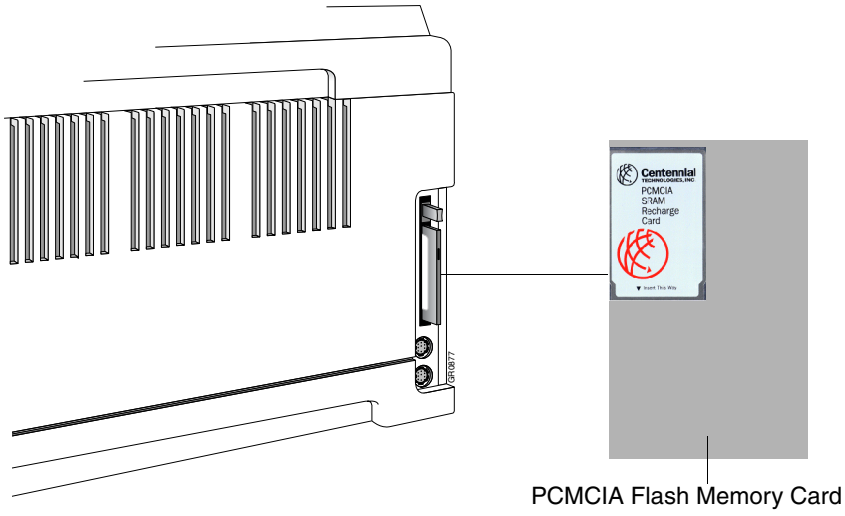
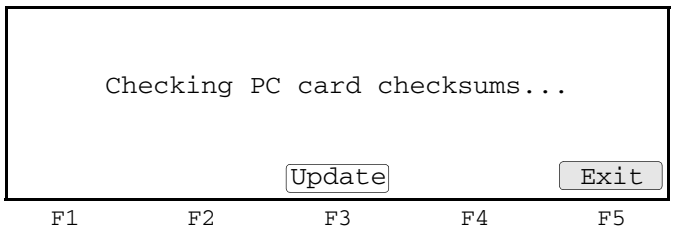
<table>
<tr><th>Step</th><th>Action</th></tr>
<tr><td>1</td><td>From the Program Manager, click the Terminal program icon.</td></tr>
<tr><td>2</td><td>From the Settings menu, choose Communication. A dialog box appears.</td></tr>
<tr><td>3</td><td>Enter the following information in the dialog box and click OK.
<table>
<tr><th>In this field...</th><th>Enter...</th></tr>
<tr><td>Baud Rate</td><td>9600</td></tr>
<tr><td>Data Bits</td><td>8</td></tr>
<tr><td>Stop Bits</td><td>1</td></tr>
<tr><td>Parity</td><td>None</td></tr>
<tr><td>Flow Control</td><td>None</td></tr>
<tr><td>Connector</td><td>Com1 or Com2 (depending on the port to which the cable is attached).</td></tr>
</table></td></tr>
<tr><td>4</td><td>From the Binary Transfers menu, choose Settings. A dialog box appears.</td></tr>
<tr><td>5</td><td>Select Kermit and click OK.</td></tr>
<tr><td>6</td><td>From the Send Binary File menu, choose Transfers.
A directory dialog box appears.</td></tr>
<tr><td>7</td><td>From the directory dialog box, find and select the 9700 upgrade file, and click OK.
A message at the bottom of the window indicates that the program is sending the file and a progress bar appears.
When the download is complete the GeneAmp PCR System 9700 automatically resets.</td></tr>
<tr><td>8</td><td>When the download is complete, from the File menu, choose Save to save these parameters.</td></tr>
</table>

Upgrading Firmware from a PCMCIA Flash Memory Card

The following procedure describes how to upgrade the firmware using a PCMCIA Flash Memory Card.

Note Contact your local Applied Biosystems representative to obtain a PCMCIA Flash Memory Card.

To upgrade using a PCMCIA Flash Memory Card:

Step	Action
1	Insert a PCMCIA Flash Memory Card that contains the updated firmware into the slot at the rear of the right side of the instrument. **Note** The label and arrow on the card face forward and toward the instrument. 
2	Press the card firmly to ensure the card is seated in the slot. **IMPORTANT** Do not remove the card or turn off the instrument until the upgrade is completed.
3	From the **Diagnostics** screen, press **Upgrade**.
4	From the **Diagnostics** screen, press **PCcard**.
5	The **Checksum** screen appears while the instrument checks the inserted PCMCIA Flash Memory Card to ensure that it contains a valid firmware upgrade. 

To upgrade using a PCMCIA Flash Memory Card: *(continued)*

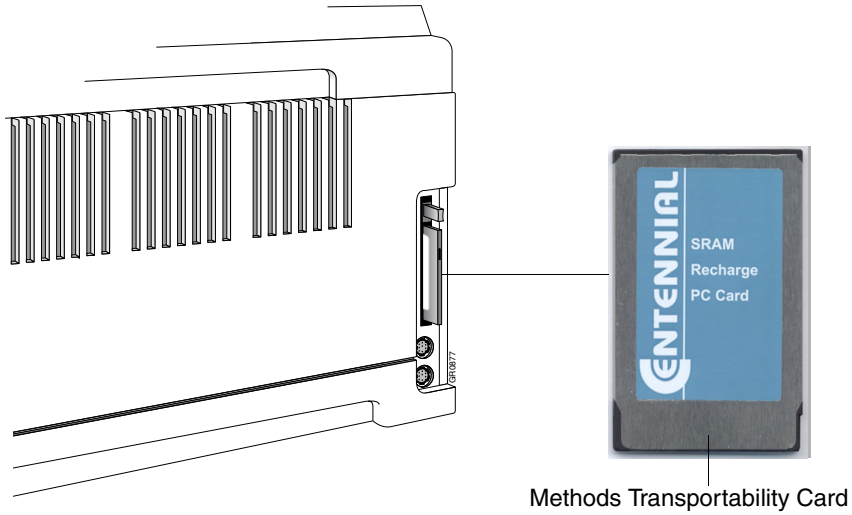
Step	Action
6	If the PCMCIA Flash Memory Card contains a valid firmware upgrade, the **Upgrade** screen appears. DO NOT TURN OFF THE INSTRUMENT OR REMOVE THE PC CARD! Upgrading the instrument firmware will take approximately 30 seconds. Instrument will re-boot when complete. F1 F2 F3 F4 F5 If successful, the **Firmware Upgrade Successful** screen displays the firmware version to which you have upgraded. Pressing the **Run** key in the lower right corner brings up the **Main** menu with the new revision number of the software listed on the screen.
7	Remove the PCMCIA Flash Memory Card after you have successfully upgraded the firmware.

Copying Methods

How to Copy Methods

You can copy methods from a Methods Transportability card to the instrument or from the instrument to a Methods Transportability card (P/N 940-1064). Runs can be made directly from methods on a Methods Transportability card.

To copy a method:

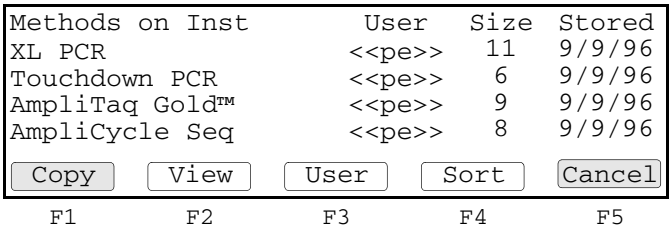
Step	Action
1	Insert a Methods Transportability card that contains the updated firmware into the slot at the rear of the right side of the instrument. **Note** The label and arrow on the card face forward and toward the instrument. 
2	Press the card firmly to ensure the card is seated in the slot. **IMPORTANT** Do not remove the card or turn off the instrument until the upgrade is completed.
3	From the **Utilities** screen, press **More**, and from this secondary utility screen, press **Copy**. The **Copy Method** dialog box appears. Copy Method ToInst - Copy a method to the instrument ToCard- Copy a method to the PC card [ToInst] [ToCard] [Cancel] F1 F2 F3 F4 F5

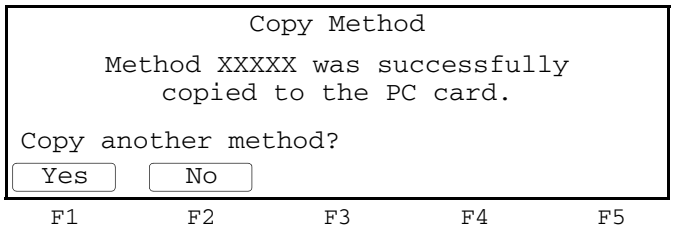
To copy a method: *(continued)*

<table>
<tr><th>Step</th><th>Action</th></tr>
<tr><td>4</td><td>In the Copy Method dialog box, you can take the following action:

<table>
<tr><th>Press...</th><th>To...</th></tr>
<tr><td>ToInst</td><td>Transfer a method from a Methods Transportability card to the instrument.</td></tr>
<tr><td>ToCard</td><td>Transfer a method from the instrument to a Methods Transportability card.</td></tr>
</table>

A screen appears with a list of the methods on the instrument or the Methods Transportability card, depending on whether you selected ToInst or ToCard.


</td></tr>
<tr><td>5</td><td>Use the Circular Key Pad to select a method and press Copy.</td></tr>
<tr><td>6</td><td>Confirmation appears when the method has been successfully copied.


</td></tr>
</table>

Connecting and Configuring a Printer

Connecting a Printer

If you connect an optional printer to your 9700 instrument, you can print out a hard copy of the time and temperature parameters for the PCR methods you create.

Connect one end of your printer cable (N805-1326) to the RS-485 serial port on the side panel of the 9700 instrument and connect the other end to the RS-232C interface serial adapter on the rear panel of the printer.

Configuring the Printer

After you have connected the printer cable to the printer, you must configure the instrument for the printer. You can connect the 9700 instrument to any printer with a serial board and the following specifications:

Baud Rate	9600
Parity	NONE
Data Bits	8
Stop Bits	1

See your printer manual for instructions on how to complete any other necessary installation steps.

4 Running PCR Samples

Overview

About This Chapter

This chapter provides information on selecting a method, starting and stopping a run, reviewing the history of a run, and what to do when the run is completed.

In This Chapter

The following topics are covered in this chapter:

Topic	See page
Selecting a Method	4-2
Running a Method	4-6
Reviewing the History of a Run	4-10
When a Run Completes	4-12

Note For information on loading and unloading samples, see the instructions accompanying your interchangeable sample block module.

Selecting a Method

Introduction

After you have prepared your samples and loaded them in the sample block, you can run a PCR amplification with a new or a stored method.

What Is a Method

A method is a set of instructions in which you specify how the instrument should heat and cool your samples in a PCR thermal profile.

Methods are stored in the instrument software.

Predefined Methods

The GeneAmp® PCR System 9700 supplies eight predefined methods that you can run:

- AmpliCycle® Sequencing
- AmpliTaq Gold® DNA Polymerase
- BigDye® Terminators
- General PCR
- LSM2
- Time Release PCR
- Touchdown PCR
- XL PCR

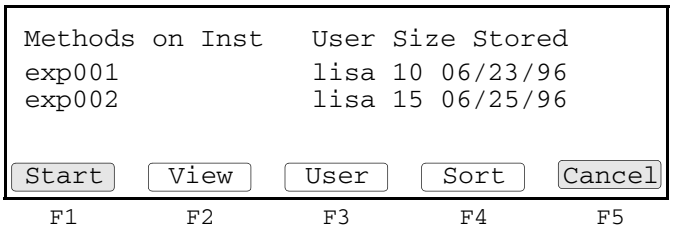
Each of these methods is stored under the user name <<pe>>. You can edit these methods and store them under a different name, a different user name, or select any one and run it.

For a detailed description of each of these pre-coded methods, and how you can use them, see Appendix B, “Supplied Methods.”

Selecting a Method

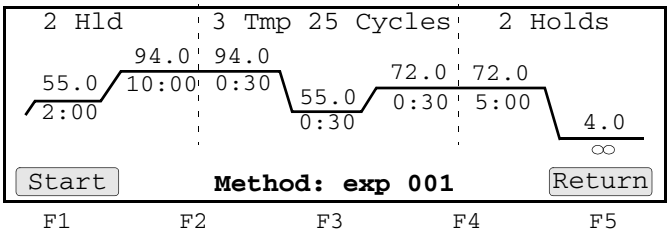
If the method you want to run has already been created and stored, you can select it from a list. If the method you want to run has not been created, see Chapter 5, “Creating and Editing Methods.”

To select a method:

Step	Action
1	From the **Main** menu, press **Run**. **Note** If a PC card is inserted, choose whether to run the method from the PC card or the instrument. The **Stored Methods** screen appears.  **Note** **Stored** represents the date the method was last saved. In the appropriate case, this column designates the date last used.
2	The units for the **Size** field are based on a calculation of the complexity and length of a method relative to a maximum size of 1102 size segments for the storage capacity of the instrument. If you need help deciding which method to select you can: ♦ View method parameters. ♦ Sort methods by different categories. ♦ Search for a method by user name.
3	Select a method by using the Circular Key Pad to move the highlight box to a method listed on the **Stored Methods** screen. **Note** You can use the up and down keys as repeat keys for quick scrolling.
4	The top line of the display continuously cycles between the following three lines: ♦ Methods on Inst User Size Stored. ♦ Used Mem: xxx methods xxx segments. **Note** The **Used Mem** field displays the number of size segments used by all stored methods. ♦ Free Mem: xxx methods xxx segments. **Note** The **Free Mem** field displays the number of size segments available to store created methods.
5	Press **Start** and start running your samples (see “Running a Method” on page 4-6).

Viewing Method Parameters

To view parameters of a method before running:

Step	Action
1	From the **Stored Methods** screen, press **View**. The **View Method** screen appears. The screen shows all the parameters of the method you selected. 
2	After reviewing PCR and post-PCR parameters of a stored method, you can: ♦ Press **Start** to start the method. ♦ Press **Return** and return to the **Stored Methods** screen. **Note** You cannot edit parameters from the **View Method** screen.

Searching for Methods

You can find any method that has been stored under a user name.

To search for a method:

Step	Action
1	From the **Stored Methods** screen, press **User**. Select which user's methods to view by taking the following action. **If you want to...** / **Then...** list all the methods currently stored on the instrument — Press All. display the methods stored under that user's name — Press Accept. **Note** You cannot add, delete, or modify a user name from this screen.
2	Making a selection returns you to the **Stored Methods** screen which now displays the methods of the user you selected.

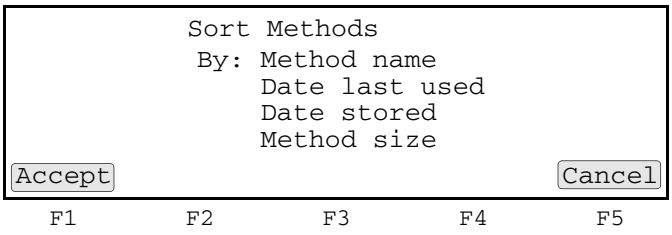
Sorting Methods

If you have a large number of stored methods, you can sort them by name, date last used, date stored and size.

To sort methods:

<table>
<tr><th>Step</th><th>Action</th></tr>
<tr><td>1</td><td>From the Stored Methods screen, press Sort.

The sorting criteria screen appears.

 
</td></tr>
<tr><td>2</td><td>Use the up and down Circular Key Pad to select the type of sort.

The following table describes the sort methods:

<table>
<tr><th>Choose this item...</th><th>To sort...</th></tr>
<tr><td>Method name</td><td>Methods alphabetically.</td></tr>
<tr><td>Date last used</td><td>Methods chronologically in descending order by date of use.

The last method which ran or was stored is listed first.</td></tr>
<tr><td>Date stored

Note Uses the most recent title and date, between date last used and the data stored.</td><td>Methods chronologically by date stored.

The last method stored is listed first.</td></tr>
<tr><td>Method size</td><td>Methods in increasing order by the amount of memory used to store each method.

The largest size method is listed first.</td></tr>
</table></td></tr>
<tr><td>3</td><td>Press Accept to accept a selection.

This returns you to the Stored Methods screen where the displayed methods are sorted according to your selection in step 2.</td></tr>
</table>

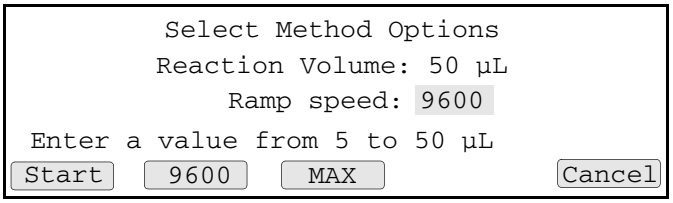
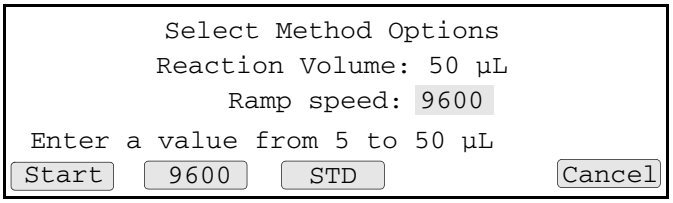
Running a Method

Starting a Run

After choosing a method, follow these steps to start a run.

To start a run:

<table>
<tr><th>Step</th><th>Action</th></tr>
<tr><td>1</td><td>From the Main menu, press Run to display stored methods.</td></tr>
<tr><td>2</td><td>Use the up and down Circular Key Pad to select the method.</td></tr>
<tr><td>3</td><td>a. Press Start.
The Select Method Options screen appears.
b. Choose from the following options:

<table>
<tr><td>96-Well Gold Sample Block Module</td><td>Select 9600 or MAX mode first, and then select volume.
 
</td></tr>
<tr><td>96-Well Aluminum Sample Block Module</td><td>Select 9600 or STD mode first, and then select volume.
 
</td></tr>
</table></td></tr>
<tr><td>4</td><td>If the displayed reaction volume differs from your reaction volume, enter the volume of your reactions in the Reaction Volume field.

The reaction volume range is dependent on the mode selected.

<table>
<tr><th>For...</th><th>The range is...</th></tr>
<tr><td>9600 mode</td><td>5 to 50 µL.</td></tr>
<tr><td>MAX/STD mode</td><td>5 to 100 µL.</td></tr>
</table>
Note Each method remembers the reaction volume.

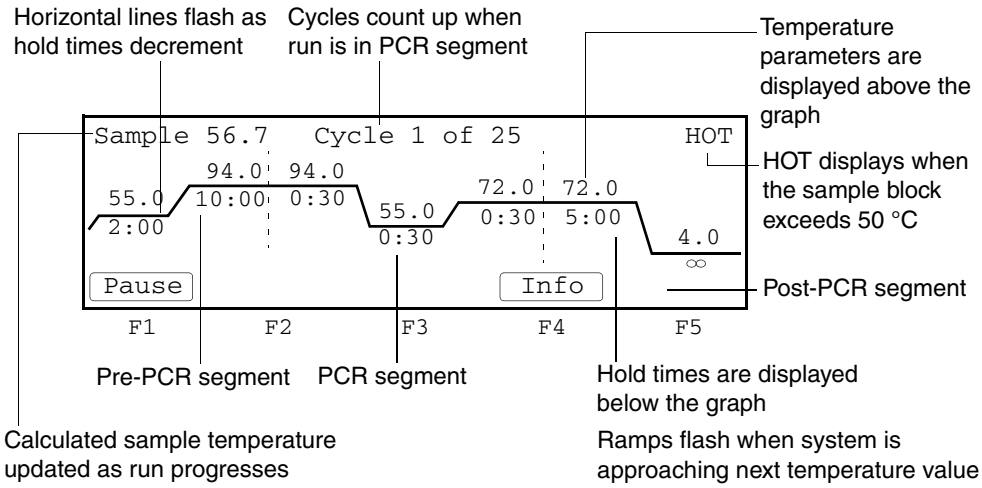
Press the CE key to clear an entry.</td></tr>
</table>

To start a run: *(continued)*

Step	Action
5	Press **Start** to start a run. If the temperature of the heated cover is less than 103 °C, this message, “Cover is heating” appears. Please wait. Cover is heating Current temperature: 65°C The run will begin when the heated cover reaches 103°C. Cancel F1 F2 F3 F4 F5
6	When the heated cover reaches 103 °C, the **Run Time** screen displays and the method you selected starts running. For a description of this screen, see “About the Run Time Screen” on page 4-7. Sample 56.7 Cycle 1 of 25 HOT 55.0 2:00 · 94.0 10:00 · 94.0 0:30 · 55.0 0:30 · 72.0 0:30 · 72.0 5:00 · 4.0 ∞ Pause Info F1 F2 F3 F4 F5

About the Run Time Screen

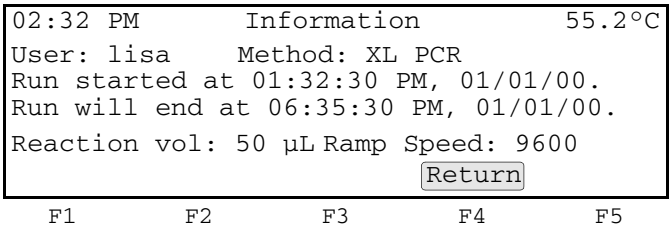
You can use the Run Time screen to chart progress at any time during the run. The Run Time screen displays the executing segment, and the next segment to execute.



Use the Run Time screen for...	For more information, see page
Viewing Method Information	4-8
Pausing a Run	4-8
Stopping a Run Before It Completes	4-9

Viewing Method Information

You can view the Method Information screen during a run by pressing Info. Press Return to return to the Run Time screen.



Pausing a Run

You can manually pause a run for a ten minute period of time during a run by pressing Pause (Figure 4-1). If you want to specify a different period of time for a pause, see “Setting the Pause Time Out” on page 3-9.

Press Resume to resume running a method before a pause expires.

Note You can pause a run in order to add a reagent. Do not to touch the sample block or the bottom of the heated cover during a pause.

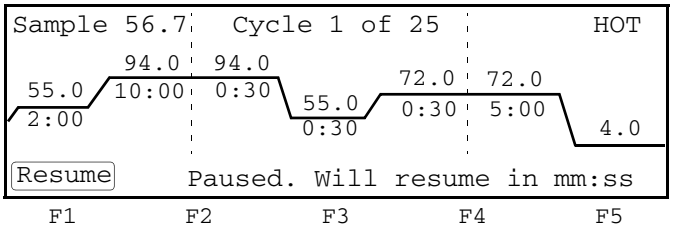


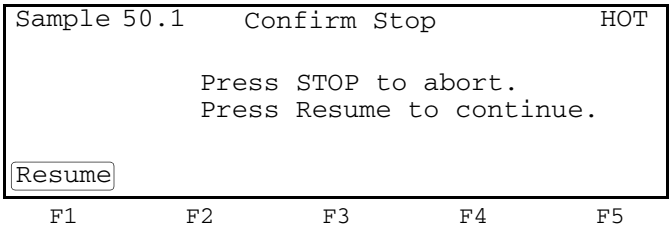
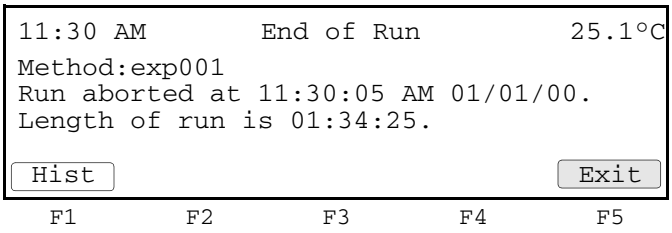
Figure 4-1 Pause screen

Your samples remain at the temperature of the instrument when you pause the run. The time remaining in a pause appears at the bottom of the screen in minutes:seconds format. It decrements to zero, and the paused run resumes at the point where you paused it.

Stopping a Run Before It Completes

The following procedure describes how to stop a run before it completes.

To stop a run before it completes:

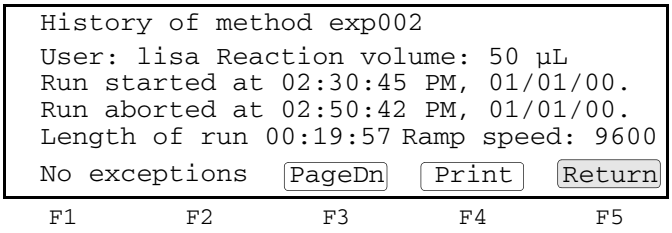
Step	Action
1	Press the **Stop** key. The **Stop confirmation** screen appears.  The run pauses for a pre-programmed period of time. When the pause time expires, the run aborts. Press **Resume** to resume. **Note** The stop and pause times are configured by setting the **Pause Time Out**. See “Setting the Pause Time Out” on page 3-9.
2	Press the **Stop** key again. This stops the run and the **End of Run** screen appears.  If any errors occur during a run the following message appears, “Exception occurred, check history file.” Press **HIST** to review the history file.
3	Press **Exit** to return to the **Main** menu.

Reviewing the History of a Run

How to Review the History of a Run

You can read a record of the events and errors that occurred during a run by reviewing the history file. The instrument stores the history file until it is overwritten by the next method used.

To review the history:

Step	Action
1	To display the History File screen: ♦ From the **Utilities** screen, press **More**, and then press **Hist**. ♦ From the **End of Run** screen, press **Hist**. 
2	Press **PageUp** to move up through the record, or **PageDn** to move down.
3	Press **Print** to print the record.

History Formats

The following table lists the history line formats.

Pre-PCR hold	<Exception> in Pre-PCR xx Setpt xx
PCR segment	<Exception> in Cycle xx Setpt xx Repxx
Any other hold	<Exception> in Hold xx Setpt xx

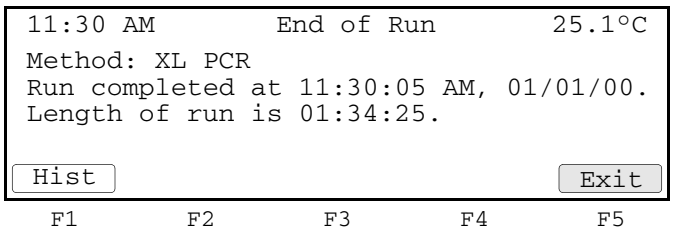
History File Records The following table lists the history file records.

Record	Description	Type
History of method xxxxxxxxxxxxxxx User xxxxxx Reaction volume xxx µL Run started at hh:mm:ss am mm/dd/yy Run ended at hh:mm:ss am mm/dd/yy Length of run hh:mm:ss RampSpeed: 9600	This header record is always created. **Note** If you stopped a method before it completed running, then the message, Run ended at..., will be Run aborted at...	Report
Power failure in Cycle xx at Setpt xx. Power failed at hh:mm:ss am for hh:mm:ss. Run resumed at hh:mm:ss am	There was a power failure during a specified point in a cycle. The message, for >18, indicates that the power was off for more than 18 hours.	Report
Drift error in Cycle xx Setpt xx Repxx. Temperature drifted x.x°c from setpt	Block drift error. The block has drifted ± 2 °C from set point during the hold segment of a run.	Report
Cover error in Cycle xx Setpt xx Repxx. Heated cover at xx.x°c	Heated cover drift error. The cover has drifted ± 5 °C from 105 °C anytime during the run.	Report
Sensor error in Cycle xx Setpt xx Repxx. Block sensor failure.	Block sensor failure.	Fatal error. Call for service.
Sensor error in Cycle xx Setpt xx Repxx. Cover sensor failure.	Heated cover sensor failure.	Fatal error. Call for service.
Setpt error in Cycle xx Setpt xx Repxx. Could not reach xx.x in hh:mm:ss	This setpoint error is only logged for setpoints above 15 °C. The limit is 5 times the normal ramping time.	Fatal error. Call for service.
Program pause in Cycle xx Setpt xx Rep xxx Method paused at xx °C for hh:mm:ss	A programmed pause was encountered.	Report
Manual pause in Cycle xx Setpt xx Rep xxx Method paused at xx °C for hh:mm:ss	You paused the run.	Report

When a Run Completes

End of Run Screen

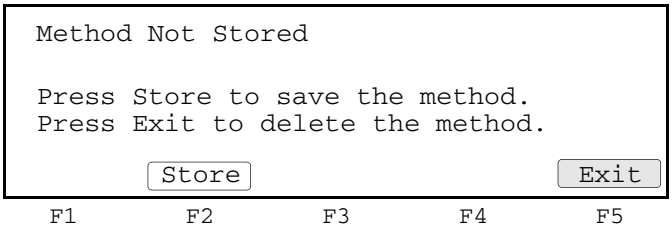
At completion of a run, the instrument beeps and the End of Run screen appears. From the End of Run screen you can perform the same functions as you can from the Stop Run screen.



If you have not yet stored the method, you must store it before exiting, or you will lose the settings. The Store soft key appears if the method has not yet been stored.

Method Not Stored Screen

If you attempt to exit the End of Run screen before storing the method, the Method Not Stored screen displays.



The following table lists the actions you can take.

If you...	Then press...
want to store the method	**Store**.
do not want to store the method	**Exit**. **Note** The **Method Not Stored** screen appears for a few seconds before the Exit key is recognized. This prevents you from losing a newly created method. You return to the top level screen and the method you created is not saved.

Creating and Editing Methods

5

Overview

About This Chapter

This chapter provides information about how to create and edit PCR methods and how to work with stored methods.

In This Chapter

The following topics are covered in this chapter:

Topic	See page
Adding or Changing Users	5-2
Creating Methods	5-6
Modifying Cycling	5-11
Printing a Method	5-17
Editing or Deleting Methods	5-18

Adding or Changing Users

Introduction

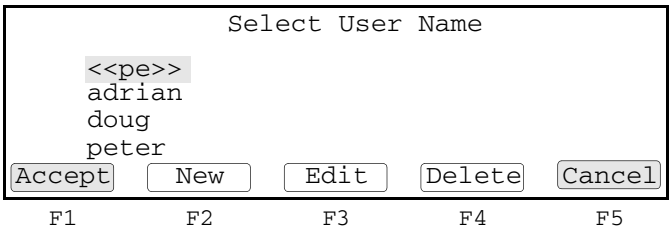
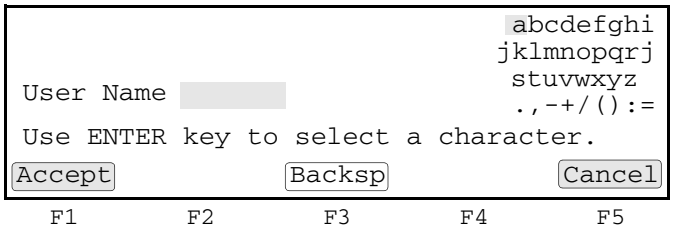
The GeneAmp® PCR System 9700 stores methods by user's names. You can add up to 19 different user names to the instrument. Once you've added your name to a list of users, and stored a method under that name, you can run the method at any time by selecting it from the Stored Methods screen (See "Selecting a Method" on page 4-3).

Use the User function on the Main menu to add new users or edit existing user names. The name you add or the name you select from a list of existing user names becomes the current user name. All new methods that you create are stored by default under the current user name.

Adding a New User Name

You add a new user name by entering an alphanumeric name on the User Name screen.

To add a new user name:

Step	Action
1	From the **Main** menu, press **User**. The **Select User Name** screen appears. A list of names of all users who have been added to the instrument displays in a 4 x 5 matrix. 
2	Press **New** to add a new name to the above list. The **User Name** screen appears. **Note** The blank space after the z is used to insert the blank spaces. 

To add a new user name: *(continued)*

<table>
<tr><th>Step</th><th>Action</th></tr>
<tr><td>3</td><td>In the User Name field, enter an alphanumeric name up to six characters in length.

You can take the following action:

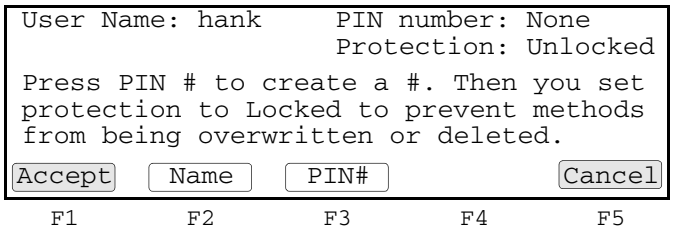
<table>
<tr><th>If you want to...</th><th>Then...</th></tr>
<tr><td>select a character in the list shown in the upper right portions of the screen</td><td>use the Circular Key Pads.</td></tr>
<tr><td>put the alphabetic character in the User Name field</td><td>press Enter.</td></tr>
<tr><td>enter the numbers directly into the User Name field</td><td>use the numeric keys.</td></tr>
<tr><td>go back one space and remove a single character</td><td>hold down the Soft key and go back multiple spaces.</td></tr>
</table></td></tr>
<tr><td>4</td><td>Press Accept to accept a name. The Security Code screen appears.

You can take the following action:

<table>
<tr><th>If you...</th><th>Then...</th></tr>
<tr><td>do not want to protect your method</td><td>press Accept again.</td></tr>
<tr><td>want to protect your method</td><td>see the following section titled “Protecting Methods.”</td></tr>
<tr><td>want to return to the Main menu without adding the new user name</td><td>press Cancel.</td></tr>
</table>
Note An error message appears if you enter a name that already exists.</td></tr>
</table>

Protecting Methods

You can protect methods and prevent other users from accidentally overwriting or deleting them by entering a Personal Identification Number (PIN#) on the Security Code screen.

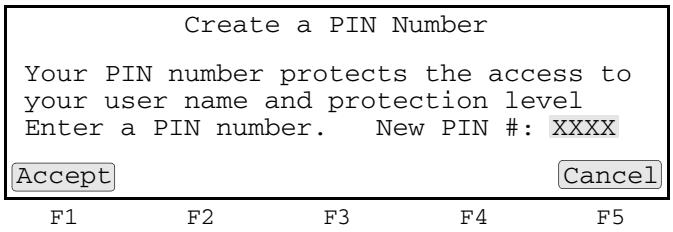
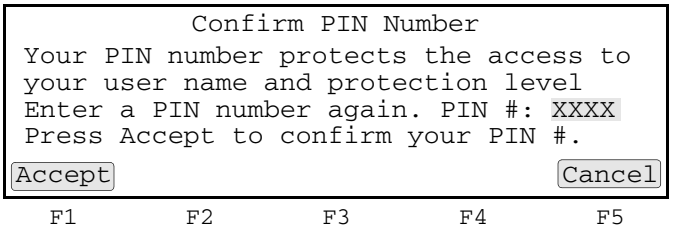
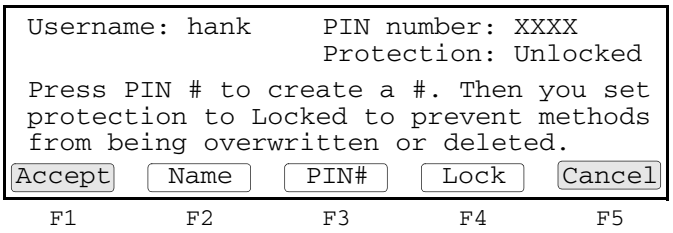


The following table lists the two levels of protection.

If a...	Then other users cannot...
user has entered a PIN #	edit that user’s name without knowing the PIN #.
method is locked	delete/overwrite the method.

Follow the procedure below to protect a method.

To protect a method:

Step	Action
1	Press **PIN #**. The **New PIN Number** screen appears. 
2	In the **New PIN #** field, use the numeric keys and type in a four-digit PIN.
3	Press **Enter**. The **PIN Confirmation** screen appears. 
4	Confirm your PIN by typing your four-digit PIN in the Confirm PIN # field.
5	Press **Enter**. The **Protection Status** screen appears. 
6	Press **Lock** to lock your method. This toggles between a Locked and Unlocked state. The **Protection** field displays the status of the method.
7	Press **Accept** when you have entered a PIN you want to keep. The **Select User Name** screen appears. The new name you entered should now display on the screen. Press **Cancel** to cancel your entry and return to the previous screen.

Changing a User Name

If you know the personal identification number for a user name, you can use the Circular Key Pad to select that name, and change it.

To change a user name:

Step	Action
1	From the **Main** menu, press **User**. The **Select User Name** screen appears.
2	Use the Circular Key Pad to select the name you want to change.
3	Press **Edit**. If a PIN has been previously entered, the **Security Check** screen appears.
4	Type in the four-digit PIN of the user name you selected.
5	Press **Name** to enter a new user name.
6	Press the **CE** key to clear the previous name.
7	Enter a new user name.
8	Press **Accept**. The Security Code screen appears again.

Deleting a User Name

If there aren't any methods stored under a user name, you can delete that name from the Select User Name screen.

To delete a user name:

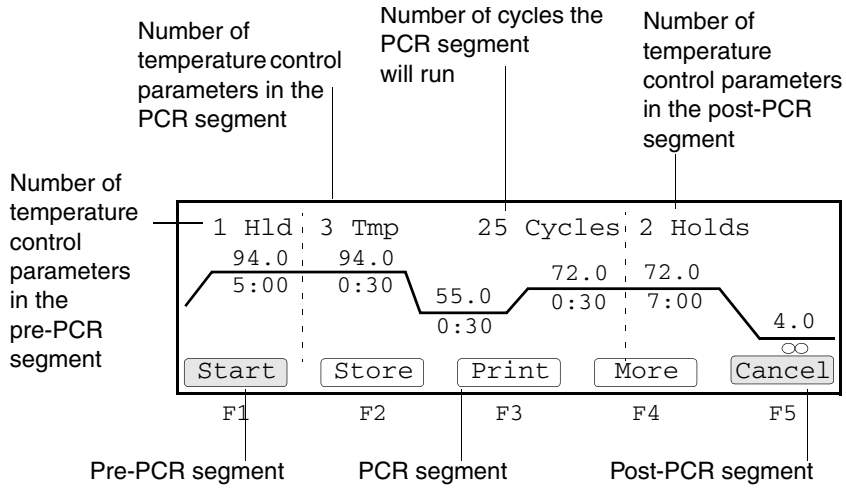
Step	Action
1	Use the Circular Key Pad from the **Select User Name** screen to select a user name.
2	Press **Delete** to delete the name. This removes the name from the Select User Name screen and allows you to add a new name to the instrument.

Creating Methods

About the Default Method

The GeneAmp PCR System 9700 comes with a default PCR thermal profile called a method. The create screen displays this default method. For information on displaying the create screen, see “Displaying the Create Methods Screen” on page 5-7.

You can run the default method shown above, or use it as a template to create a customized method.



Basic Parameters

To create a method, you need to define the following four basic parameters:

- Temperature Control Parameters
- Pre-PCR Holds
- PCR Parameters
- Post-PCR Holds

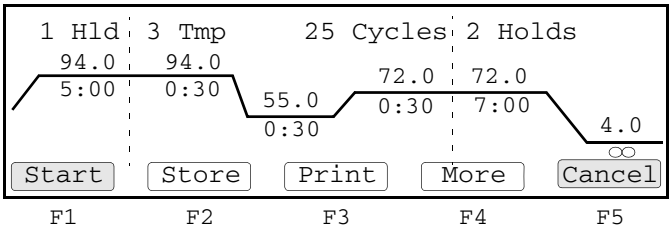
For detailed information about these parameters refer to the table below:

For information about...	See page
Entering Temperature Control Parameters	5-8
Defining Pre-PCR Holds	5-9
Defining PCR Parameters	5-9
Defining Post-PCR Holds	5-10

Displaying the Create Methods Screen

Follow the steps below to display the Create Methods screen.

To display the Create Methods screen

Step	Action
1	From the **Main** menu, press **Create**. The create screen appears. 
2	From the **Create** screen, you can take the following action:

If you want to...	Then...
start running the default method	press **Start**.
store the method under a user's name	press **Store**.
print a record of the method parameters	press **Print**.
enter temperature control parameters	enter the information on the create screen and create a new method.
display more functions for modifying methods	press **More**. **Note** The More function only displays when you select a time or temperature parameter. For detailed information on using the More function to modify cycles, see "Modifying Cycling" on page 5-11.
return to the previous screen	press **Cancel**.

Entering Temperature Control Parameters

When you enter temperature control parameters, you define values for parameters in each of the three segments of a method: pre-PCR, PCR, and post-PCR.

To enter temperature control parameters:

Step	Action
1	On the **Create** screen, select a field. When you first display the Create screen, the **Hld** field is highlighted.
2	Use the numeric keys to enter values.
3	Press **Enter** to accept a value. The next field is then selected in the order shown in Figure 5-1.

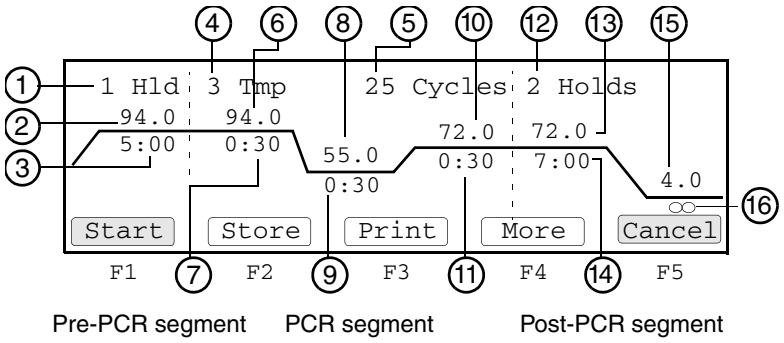


Figure 5-1 Order of advancement of the highlight box

Number	Description
1	Number of pre- PCR holds
2	Pre-PCR temperature parameter
3	Pre-PCR hold time parameter
4	Number of PCR segment temperatures
5	Number of cycles for the PCR segment
6	PCR segment temperature parameter
7	PCR segment time parameter
8	PCR segment temperature parameter
9	PCR segment time parameter
10	PCR segment temperature parameter
11	PCR segment time parameter
12	Number of post-PCR holds
13	Post-PCR temperature parameter
14	Post-PCR hold time parameter
15	Post-PCR temperature parameter
16	Post-PCR hold time parameter

Defining Pre-PCR Holds

The Hld field on the Create screen defines the number of holds for the pre-PCR segment of your method. One (1) is the typical setting for most PCR amplifications and is the default value for the Hld field.

Generally, pre-PCR holds define a temperature and hold-time sufficient to denature any endonucleases or exonucleases that may contaminate your prepared samples.

To define pre-PCR holds:

Step	Action
1	On the **Create** screen, in the **Hld** field, type in the number of pre-PCR holds for your method. **Note** You can enter 0 in this field to delete the pre-PCR hold segment from your method.
2	Create the first temperature parameter: a. Press **Enter**. b. Type in a temperature value between 4.0 °C and 99.9 °C.
3	Create the first hold-time parameter: a. Press **Enter**. b. Type in a hold-time value between 00:00 and 98:59 (minutes:seconds).
4	Enter information for the next pre-PCR hold: a. Press **Enter**. b. Repeat step 2 and step 3 until you have hold-time and temperature values for each of the pre-PCR hold parameters you defined in step 1.

Defining PCR Parameters

The Tmp field on the Create screen defines the number of temperature control parameters in the PCR cycling segment of your method. Three temperature PCR is the typical setting for many PCR amplifications:

- Template denaturation
- Primer annealing
- Primer extension

To define the PCR parameters:

Step	Action
1	On the **Create** screen, in the **Tmp** field, type in the number of temperature control parameters (2–6) you want for the PCR segment of your method. **Note** Specifying only the minimum number of PCR cycles required for analysis will minimize the chance that unwanted targets will amplify competitively.
2	Enter the number of cycles you want the method to run: a. Press **Enter** to select the **Cycles** field. b. In the **Cycles** field, type in the number of cycles (from 2–99). **Note** Twenty-five cycles is the default setting.
3	Create the first temperature parameter: a. Press **Enter**. b. Type in a temperature value between 4.0 °C and 99.9 °C.

To define the PCR parameters: *(continued)*

Step	Action
4	Create the first hold-time parameter: a. Press **Enter**. b. Type in a hold-time value between 00:00 and 98:59 (minutes:seconds).
5	Repeat step 3 and step 4 until you have time and temperature values for each of the segment temperature control parameters you defined in step 1.

Defining Post-PCR Holds

On the Create screen, the Holds field defines the number of temperature control parameters in the post-PCR segment of your method.

The post-PCR incubation temperature and hold time parameters define how to hold your samples at a specified temperature until you are ready to analyze them.

Note If the idle state setpoint, or the last hold of the Method are below 15 °C, then the heated cover will automatically set to 50 °C.

Post-PCR Parameter Settings

Typical Post-PCR parameter settings:

Temperature	Time (min:sec)	Use For
72 °C	7:00	Complete extension of all amplicons
72 °C	99:59 (×)	AmpErase™ applications
4 °C	99:59 (×)	General storage

Follow the procedure below to define Post-PCR Holds

To define post-PCR holds:

Step	Action
1	Select the **Holds** field.
2	In the Holds field, type in the number of post-PCR steps for your method.
3	Press **Enter** to select the first post-PCR temperature parameter.
4	Type in a temperature value between 4.0 °C and 99.9 °C.
5	Press **Enter** to select the first post-PCR hold time parameter.
6	Type in a hold time value between 00:00 and 98:59 (*min:sec*). **Note** The hold time × indicates a hold that lasts indefinitely. You can enter an × hold time, by typing a hold time value of 99:00 or greater.
7	Press **Enter**. This selects the next temperature parameter.
8	Repeat step 4 through step 7 until you have time and temperature values for each of the post-PCR hold parameters you defined in step 2.

Modifying Cycling

Introduction

In addition to customizing values for PCR temperature control parameters, you can use the More function on the create screen and access cycle modification functions that allow you to:

- Auto-increment/decrement time and temperature parameters.
- Modify up-ramp and down-ramp rates in the cycling segment of a method.
- Insert holds, cycles, and programmed pauses.
- Delete temperature control parameters.

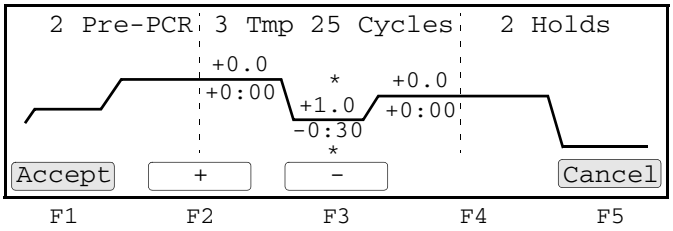
The time or temperature parameter you select on the create screen, determines which modification function you can access when you press More. Different modification functions are available depending on whether you select a temperature control parameter in the pre-PCR segment, the PCR segment, or the post-PCR segment of a method.

Changing Temperature Control Parameters

Using the AutoX function, you can automatically increase or decrease the value for any PCR segment parameter by a fixed amount every cycle.

Note This feature is particularly useful towards the end of the amplification process since the amount of PCR product, available to be extended, increases with the number of cycles while the amount of available enzyme remains constant.

To automatically change temperature control parameters:

Step	Action
1	From the **Create** screen, use the Circular Key Pad to select a time or temperature parameter in the PCR segment.
2	Press **More**. The **Modify** screen appears.
3	Press **Modify**. The **Select Modification** screen appears.
4	Press **AutoX**. The **AutoX** screen appears.  **Note** If you have inserted a programmed pause, the AutoX screen displays the pause, but you cannot modify it from the AutoX screen.
5	Select the PCR time or temperature parameter that you want modified when you run your method. **Note** From the AutoX screen, you cannot modify the number of parameters in each segment or the number of cycles.

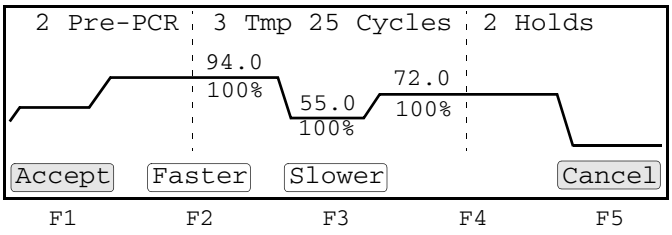
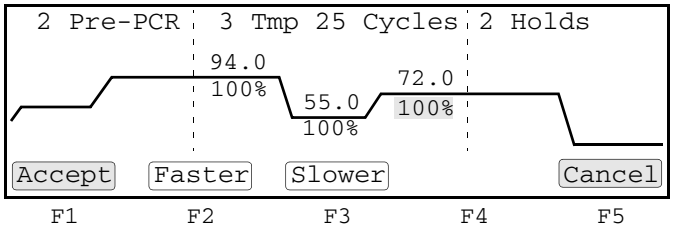
To automatically change temperature control parameters: *(continued)*

Step	Action
6	You can take the following action: **If you want to...** / **Then press...** increase the value every cycle (a plus sign displays in the current field) — + (plus sign). decrease the value every cycle (a minus sign displays in the current field) — - (minus sign). An asterisk * appears on method screens for parameters that have been modified.
7	Press **Accept** to accept all entries on the AutoX screen.

Modifying Ramp Rates

The ramp time is the time it takes the instrument to change from one temperature to another. Using functions accessible from the Modify screen, you can modify the up-ramp and down-ramp rates of the instrument by defining it as a percentage of the temperature's maximum rate of increase. The default maximum up-ramp and down-ramp rates is 100%.

To modify ramp rates:

Step	Action
1	Press **Modify** from the Modify screen. The **Select Modification** screen appears.
2	Press **Ramp**. The **Ramp Rate Modification** screen appears.  In this example, you can modify the rate at which the instrument ramps up from 72.0 °C to 94.0 °C, down from 94.0 °C to 55.0 °C, or up from 55.0 °C to 72.0 °C.
3	Use the Circular Key Pad to select a ramp that you want to modify.
4	Move the highlight box to the third temperature in the cycling segment of the default method to modify the highlighted ramp rate. The third ramp rate defines the rate at which the instrument increases from 55.0 °C to 72.0 °C each time the method cycles. 

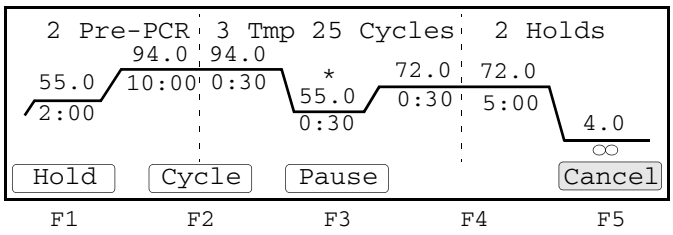
To modify ramp rates: *(continued)*

Step	Action
5	Modify the up-ramp and down-ramp rates you selected by defining them as a percentage of the maximum of 100%: **If you want to...** / **Then press...** increase the ramp rate by 10% up to a maximum of 100% / the **Faster** key. decrease the ramp rate by 10% from 100% to 10%, and by 5% from 10% to 5% / the **Slower** key. **Note** You can also use the numeric keys to enter a value that defines the percentage by which you want to decrease the ramp rate for each cycle of the method. You can enter values between 5 and 95, or 100. For ramp rates less than 100%, an asterisk * appears next to modified ramps. The asterisk remains beneath the modified temperature parameter to remind you that the method has been modified. **Note** The following message appears if you enter a numeric value outside the range of acceptable values, “Valid range is 5 to 95 and 100.”
6	Press **Accept** to accept all entries, and return to the previous screen.

Inserting Holds

Use the Insert function to insert holds and cycles into your method, and program pauses that the instrument automatically inserts into your method as it runs.

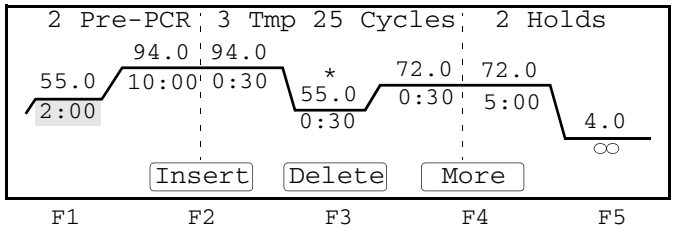
To insert holds:

Step	Action
1	From the **Create** screen, use the Circular Key Pad to select a time or temperature parameter to the right of where you want to insert a hold.
2	Press **More**. Depending on the parameter you select in step 1, one of three screens displays from which you can access the insert function.
3	Press **Insert**. The Insert screen appears.  **Note** The Pause function will not display on the Insert screen if the highlighted segment already has a pre-programmed pause or if highlight box is not on a PCR segment parameter.
4	Press Hold to insert a hold of 4.0 °C for 30 seconds to the left of the parameter you selected in step 1.
5	Type in a value for the hold temperature.
6	Type in a value for the hold time.
7	Press More to return to the Create screen. Your modified method now appears.

Deleting Holds

Follow the procedure below to delete holds.

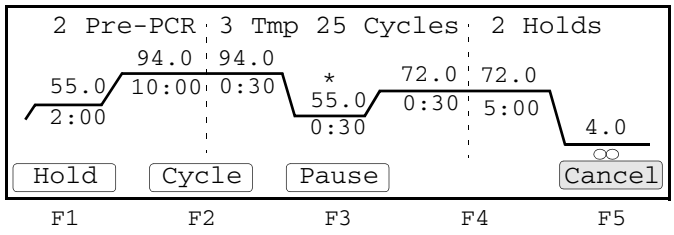
To delete holds:

1	From the **Edit** screen, select a hold parameter and press **More**.
2	Use the Circular Key Pad to a select a time or temperature hold parameter. 
3	Press **Delete**.

Inserting Cycles

Follow the procedure below to insert cycles.

To insert cycles:

Step	Action
1	From the **Create** screen, use the Circular Key Pad to select a time or temperature parameter to the right of where you want to insert a cycle.
2	Press **More**. Depending on the parameter you select in step 1, one of three screens displays from which you can access the insert function.
3	Press **Insert**. The Insert screen appears. 
4	Press **Cycle** to insert a cycle to the left of the segment you selected in step 2. **Note** You can delete a cycle by entering 0 in the **Tmp** field.

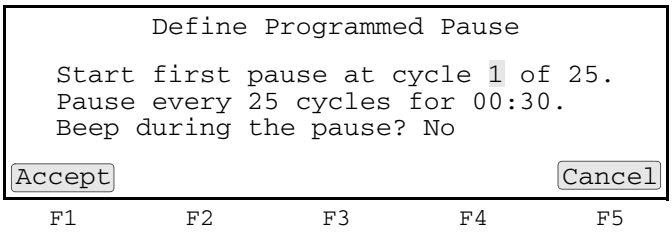
Inserting Programmed Pauses

Follow the procedure below to insert a programmed pause.

To insert a programmed pause:

Step	Action
1	From the **Create** screen, use the Circular Key Pad to select a PCR segment time or temperature parameter where you want to insert a programmed pause.
2	Press **More**.
3	Press **Insert**. The Insert screen appears.

To insert a programmed pause: *(continued)*

Step	Action
4	From the Insert screen, press **Pause** . The **Programmed Pause** screen appears. 
5	In the **Start first Pause at Cycle** field, type in the cycle number where you want the method to first pause.
6	In the **Pause Every** field, type in the pause frequency in cycles. The pause frequency specifies the number of cycles that will run between each pause.
7	In the **Cycles For** field, type in the length of the pause in minutes:seconds (00:01–98:59) format.
8	In the **Beep During The Pause?** field, press **Yes** or **No**.
9	Press **Accept** to accept the pause information on the screen. The word **Pause** now displays to the right of the incubation step where you programmed the pause. **Note** You can only insert one pause in each cycle.

Editing Programmed Pauses

If you have inserted a programmed pause in your method, you can edit the parameters for the pause at any time.

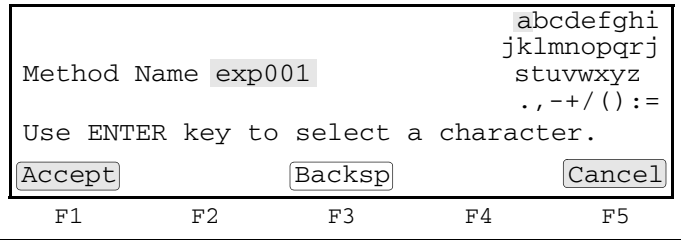
To edit programmed pauses:

Step	Action
1	From the **Create** screen, use the Circular Key Pad to highlight the word **Pause**. The **Edit** soft key appears.
2	Press **Edit** to access the programmed pause screen. From this screen, you can ♦ Change any of the pause parameters, or ♦ Use the Circular Key Pad to select the pause time parameter on the screen, and edit it by entering a different time. After you have entered all modifications to the customized method you are creating, you should store the method before running it (see "Naming and Storing Methods" below).

Naming and Storing Methods

Naming and storing completes the creation of the method.

To name and store methods:

Step	Action
1	From the **Create** screen, press **Store**.
2	You can take the following action: **If you want to...** / **Then...** store the method under the name displayed in the user field and name the method the default name displayed in the method field The default method name is expxxx where xxx= a number from 0 to 999. — press the **Accept** key. rename the method — go to step 3.
3	From the **Store** screen, press **Method**. The Method Name screen appears. **Note** The blank space after the letter Z is used to insert blank spaces. 

To name and store methods: *(continued)*

Step	Action
4	In the **Method Name** field, follow these steps to enter a 1–16 character alphanumeric name. **Step** / **Action** a. Use the Circular Key Pad to select a character in the list shown in the upper right portion of the screen. b. After selecting a character, press **Enter** to place the character in the **Method Name** field. c. **If you want to...** / **Then...** enter a number — press the appropriate number key. go back one space — press the **Backsp** key. clear the method name — press the **CE** key.
5	Press **Accept** after you have entered a method name. The **Store** screen appears again. The method name you entered should now display in the Method Name field.
6	From the **Store** screen, press **Accept**. This stores the method under the name you entered.

Printing a Method

Introduction

If you have configured your instrument for a printer, you can print a record of the parameters in a method. For more information on configuring a printer, see “Setting Custom Parameters” on page 3-8.

Printing a Method

To print a method:

Step	Action
1	Access the **Create** or **Edit** screen. From the **Create** or **Edit** screen you can print a copy of the parameters for the method displayed on the screen.
2	Press **Print**.

Editing or Deleting Methods

Introduction

After you create a method, you can edit its parameters, and store the method by the same name, or change its name. At some time, you may also want to delete a method if you are no longer using it. You can access all editing functions from the Main menu. The delete screen is accessed through the utility menu.

Editing a Method

The following procedure describes how to edit a method.

To edit a method:

<table>
<tr><th>Step</th><th>Action</th></tr>
<tr><td>1</td><td>From the Main menu, press Edit.

Note If an (SRAM) PC Card is detected in the Card slot, you will get a choice of editing a method on the PC Card or the instrument.

The top line of the display continuously cycles between the following three lines:
Methods on Inst User Size Stored [or Last Used]
[or on PC card]
Used Mem: xxx methods xxx segments
Free Mem: xxx methods xxx segments

The following table describes these fields.

<table>
<tr><th>Field</th><th>Description</th></tr>
<tr><td>units for the Size field</td><td>Based on a calculation of the complexity and length of a method relative to a maximum size of 1102 size segments for the storage capacity of the instrument.</td></tr>
<tr><td>Used Mem field</td><td>Displays the number of segments used by all stored methods.</td></tr>
<tr><td>Free Mem field</td><td>Displays the number of segments available to store created methods.</td></tr>
</table>
</td></tr>
<tr><td>2</td><td>Select one of the methods displayed on the screen, or select another method as follows:

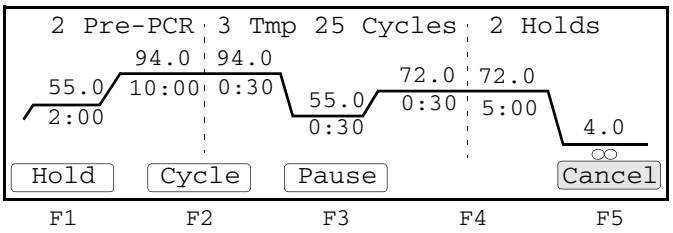
<table>
<tr><th>If you want to...</th><th>Then press...</th></tr>
<tr><td>view the parameters of a method before making a selection

(Refer to “Viewing Method Parameters” on page 4-4.)</td><td>the View key.</td></tr>
<tr><td>search for a method by user name

(Refer to “Searching for Methods” on page 4-4.)</td><td>the User key.</td></tr>
<tr><td>sort methods by different criteria

(Refer to “Sorting Methods” on page 4-5.)</td><td>the Sort key.</td></tr>
</table>
</td></tr>
</table>

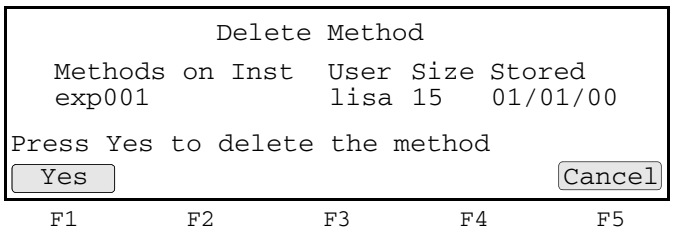
To edit a method: *(continued)*

Step	Action
3	Press **Edit** after selecting a method. The Edit screen appears. 
4	Choose a temperature or time parameter within a PCR segment.
5	Edit temperature control or time parameters. **Note** Editing parameters on the Create screen involves the same tasks and uses the same key combinations as you use when creating a method. The same functions for modifying methods are also available.
6	From the **Edit** screen, press **Store** to store the method.

Deleting a Method

The following procedure describes how to delete a method.

To delete a method:

Step	Action
1	From the **Utilities** screen, press **More** and then press **Delete**. The Delete Method screen appears.  **Note** If a PC Card is detected in the Card slot, you will get a choice of deleting a method on the PC Card or the instrument. The top line of the display continuously cycles between the following three lines: Methods on Inst [or on PC card] User Size Stored [or Last Used] Used Mem: xxx methods xxx segments Free Mem: xxx methods xxx segments The following table describes these fields. **Field** / **Description** Units for the Size field — Based on a calculation of the complexity and length of a method relative to a maximum size of 1102 size segments for the storage capacity of the instrument. Used Mem Field — Displays the number of segments used by all stored methods. Free Mem Field — Displays the number of segments available to store created methods.
2	Select one of the methods displayed on the screen, or select another method as follows: **If you want to...** / **Then press...** view method parameters (Refer to “Viewing Method Information” on page 4-8.) — the **View** key. sort methods by different criteria (Refer to “Sorting Methods” on page 4-5.) — the **Sort** key.
3	Press **Delete**. The Delete Confirmation screen appears. If the method is protected, enter a four-digit PIN and press Accept when the number is correct.

To delete a method: *(continued)*

Step	Action
4	Press **Yes** to confirm the deletion. This deletes the method and returns you to the Delete screen. **Note** Even after you delete the last method stored under a User name, the name is removed from the instrument. To delete the name, see “Deleting a User Name” on page 5-5.

6 Converting Hold Times

Overview

About This Chapter

This chapter provides information about setting hold times for the GeneAmp® PCR System 9700 compared to the DNA Thermal Cycler or DNA Thermal Cycler 480. This information is based upon differences in how the instruments heat and cool samples.

Note Protocols using sample volumes between 5 µL and 50 µL, developed on either the GeneAmp® PCR System 2400 or 9600, may be transported to the GeneAmp PCR System 9700 without change when using the 9600 emulation mode.

In This Chapter

The following topics are covered in this chapter:

Topic	See page
About Setting Hold Times	6-2
Guidelines for Converting Hold Times	6-3

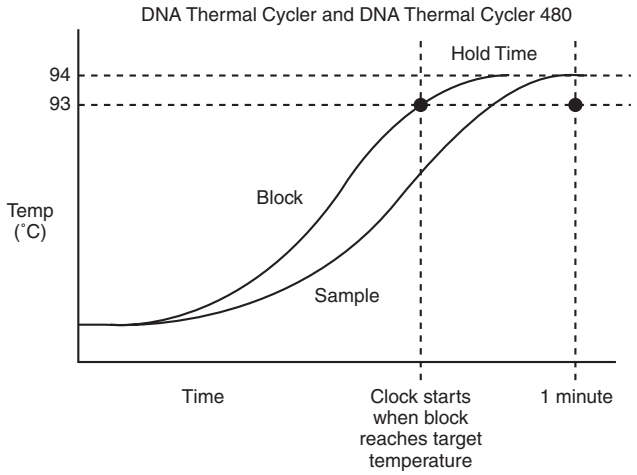
About Setting Hold Times

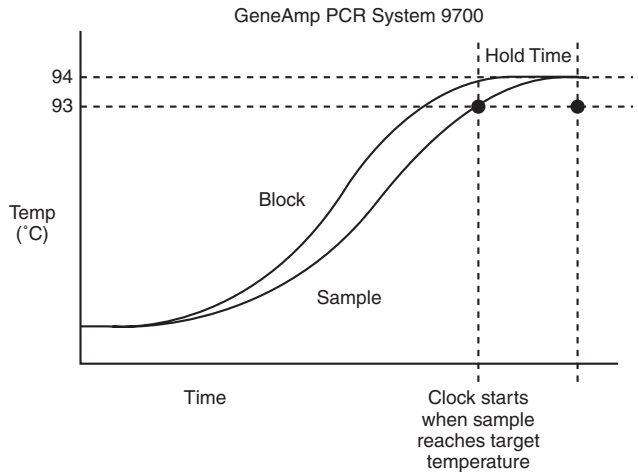
Introduction

The hold times specified for the GeneAmp PCR System 9700 are shorter than those used for the Step-Cycle or ThermoCycle files on the DNA Thermal Cycler and the DNA Thermal Cycler 480. This is because the DNA Thermal Cycler and the DNA Thermal Cycler 480 starts counting the hold time when the block reaches a temperature one degree before the target temperature, while the GeneAmp PCR System 9700 starts counting the hold time when the samples reach a temperature one degree before the target temperature.

Hold Time Differences

As shown in the figures below, since the block reaches the target temperature before the sample, the programmed hold time on the DNA Thermal Cycler and the DNA Thermal Cycler 480 must include enough time for the samples to reach the target temperature. A hold time of one minute or greater is required on the DNA Thermal Cycler and the DNA Thermal Cycler 480 for samples to reach the target temperature. On the GeneAmp PCR System 9700, hold times of less than one minute are generally used.





Guidelines for Converting Hold Times

Introduction

This section describes guidelines on how to convert hold times for the DNA Thermal Cycler or the DNA Thermal Cycler 480 to hold times for the GeneAmp PCR System 9700 when using the 9600 mode.

What to Base the New Hold Times On

The following table lists what to base the new hold time on if you are using an up ramp or down ramp.

If you are using...	Then base the new hold time...	For more information see...
an up ramp	on the change in the temperature required to reach the next target temperature.	Table 6-1 on page 6-3.
a down ramp	on the starting temperature of the ramp and the change in temperature required to reach the next temperature.	Table 6-2 on page 6-5.

Setting Up Ramps

The following table lists the process of setting the up ramp temperature.

Step	Action
1	Determine the change in temperature required to reach the next target temperature and round this value off to the closest value found in Table 6-1.
2	Subtract the number of seconds indicated from the hold time used for the DNA Thermal Cycler or the DNA Thermal Cycler 480. The result is the hold time to use for the GeneAmp PCR System 9700. **Note** The typical hold time is 10 to 15 seconds for denaturation.

Table of Conversions

The following table lists the times for converting up ramp hold times greater than one minute from the DNA Thermal Cycler and the DNA Thermal Cycler 480 to the GeneAmp PCR System 9700.

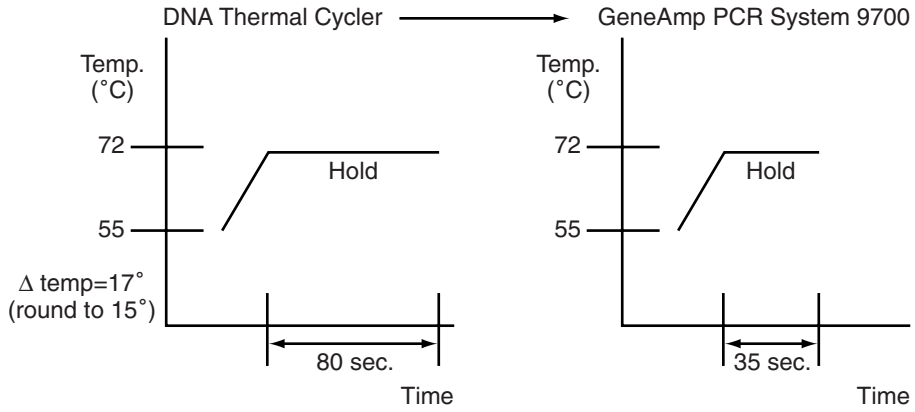
Table 6-1 Converting up ramp hold times

∆ Temp (°C)	Seconds to subtract from DNA Thermal Cycler or DNA Thermal Cycler 480 hold times (>1 min.)
10°	38 sec.
15°	45 sec.
20°	49 sec.
30°	54 sec.
40°	55 sec,
50°	57 sec.
60°	57 sec.

Up Ramp Example

In this example, the temperature was increased by 17 °C. This value was rounded to 15 °C. According to Table 6-1, subtract 45 seconds from the hold time on the DNA Thermal Cycler or the DNA Thermal Cycler 480, resulting in a new hold time of 35 seconds (see the figure below).

Note If methods developed on the GeneAmp PCR System 9700 will be used on the DNA Thermal Cycler or the DNA Thermal Cycler 480, you can also use Table 6-1 to convert the hold times. Add the indicated times instead of subtracting them.



Setting Down Ramps

The following table lists the process of setting the down ramp temperature.

Step	Action
1	Determine the change in temperature required to reach the next target temperature and round this value off to the closest value found in Table 6-2.
2	Based on the starting temperature of the ramp, determine the number of seconds to subtract from the DNA Thermal Cycler or DNA Thermal Cycler 480 hold time to arrive at the new hold time. **Note** The typical hold time is 10 to 15 seconds for annealing.

Table of Conversions

The following table lists the times for converting down ramp hold times greater than one minute from the DNA Thermal Cycler or the DNA Thermal Cycler 480 to the GeneAmp PCR System 9700.

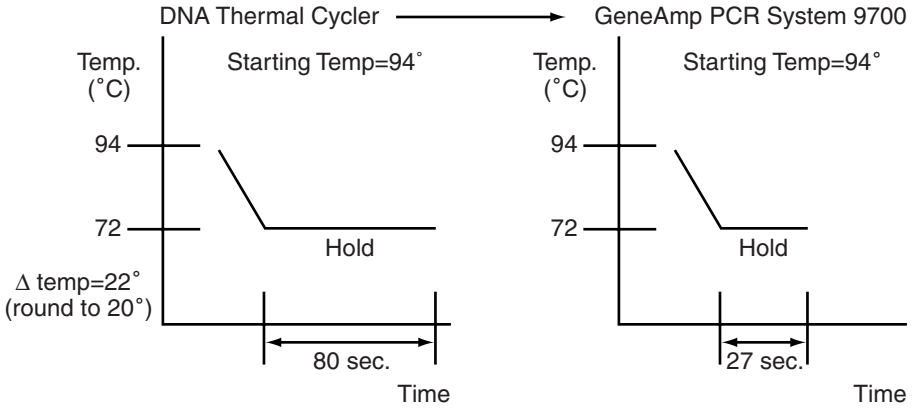
Table 6-2 Converting down ramp hold times

	Starting temperature of ramp (°C)		
A Temp (°C)	**35-55°**	**56-75°**	**76-96°**
10°	39 sec.	39 sec.	40 sec.
15°	45 sec.	47 sec.	48 sec.
20°	49 sec.	52 sec.	53 sec.
30°	—	56 sec.	59 sec.
40°	—	57 sec.	62 sec.
50°	—	—	62 sec.
60°	—	—	60 sec.

Down Ramp Example

In the example below, the temperature was decreased by 22 °C. This value was rounded to 20 °C. According to Table 6-2, with a starting temperature of 94 °C, we should subtract 53 seconds from the hold time on the DNA Thermal Cycler or the DNA Thermal Cycler 480, resulting in a new hold time of 27 seconds.

Note If the methods developed on the GeneAmp PCR System 9700 will be used on the DNA Thermal Cycler or the DNA Thermal Cycler 480, use Table 6-2 to convert the hold times. Add the indicated times instead of subtracting them.



7 Routine Maintenance

Overview

About This Chapter

This chapter describes how to perform routine maintenance on the GeneAmp® PCR System 9700.

DANGER **ELECTRICAL SHOCK HAZARD.** Severe electrical shock can result from operating the GeneAmp PCR System 9700 Base Module without its instrument panels in place. Do not remove instrument panels. High-voltage contacts are exposed when instrument panels are removed from the instrument.

There are no components inside the GeneAmp PCR System 9700 that you can safely service yourself. If you suspect a problem, contact an Applied Biosystems Technical Support Representative.

In This Chapter

The following topics are covered in this chapter:

Topic	See page
Changing the External Fuses	7-2

Changing the External Fuses

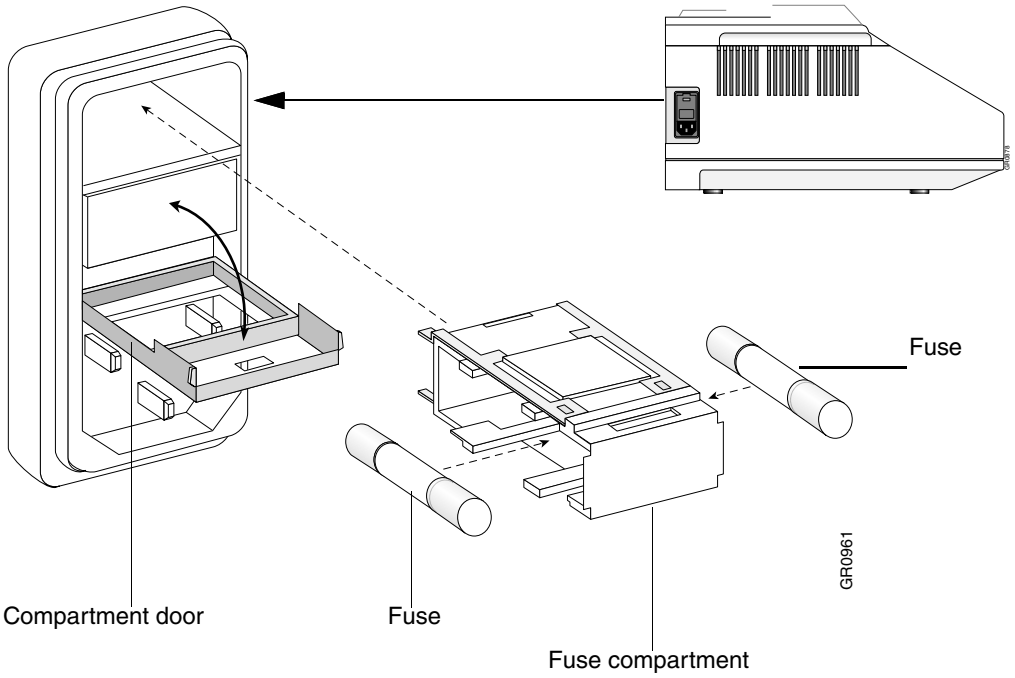
Introduction

WARNING **FIRE HAZARD**. For continued protection against the risk of fire, replace fuses only with Listed and Certified fuses of the same type and rating as those currently in the instrument.

All instruments have factory installed fuses. However, if you should ever need to change a fuse, follow the instructions in “Changing the External Fuses” on page 7-2.

Power Entry Module Diagram

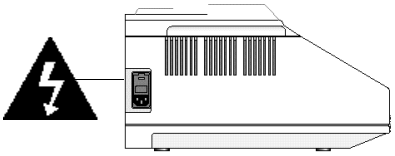
The Power Entry Module is located at the rear of the left side of the instrument.



Changing Fuses

Instruments have two 8 amp Type T 250 V 5x20 mm fuses (P/N 0999-1683).

To change the fuses:

Step	Action
1	Turn off the instrument and disconnect the power cord from the side of the instrument. **DANGER** **ELECTRICAL SHOCK HAZARD**. Severe electrical shock, which could cause physical injury or death, can result from working on an instrument when the high voltage power supply is operating. To avoid electrical shock, disconnect the power supply to the instrument, unplug the power cord, and wait at least 1 minute before working on the instrument.  The fuses are located in the Power Entry Module.
2	Insert a small flat-tip screwdriver into the slot in the upper portion of the power entry module, and open the door.
3	Pull the fuse compartment out. There are two fuses in the fuse compartment.
4	Pull out the fuse from the back of the fuse compartment and replace the blown fuse with one 8 amp Type T 250 V fuse.
5	Place the fuse compartment back into the Power Entry Module and close the door. Press the door until it locks in place.
6	Connect the instrument power cord.

8 Troubleshooting

Overview

About This Chapter

This chapter describes instrument problems you may have, the probable causes of these problems, and any display screen messages you may encounter when using the GeneAmp® PCR System 9700.

In This Chapter

The following topics are covered in this chapter:

Topic	See page
If There Is a Power Failure	8-2
Display Screen Error Messages	8-3
Troubleshooting Information	8-6

If There Is a Power Failure

Introduction

An automated restart function allows for power failures and safe continuation of a PCR run after resumption of power.

During a Power Failure

The following table lists the actions the instrument takes if the power is interrupted and the instrument turns off while you are operating it.

Note If the power is off for 15 seconds or longer and fails during execution of a cycle then the cycle currently running will restart. If the power fails while executing a hold, or approaching a hold, then that hold temperature will restart from the beginning.

Note If the power failure lasts longer than 18 hours, the Resume will not occur.

Do the following in a power failure:

Step	Action
1	Restart or continue the PCR experiment. The instrument determines what temperature was being approached, or was holding. Upon resumption of power, it will go to that temperature and countdown the time remaining in the hold as soon as the temperature is within the specified clock start limits.
2	Incubate the samples until you can continue the experiment.
3	Enter a record for any power outage in the history file.

Display Screen Error Messages

Error Messages Table Refer to the following table for a description of error messages, and recommended actions that you should take.

Table 8-1 Error Messages

Message	Description	Recommended Action
Battery RAM version number lost	This error is generated when the battery RAM has been lost and re-initialized.	Call Technical Support.
Block Calibration initialized	Software or hardware failure.	Call Technical Support.
Block Calibration reset to default	System error. Block data reset to defaults.	Call Technical Support.
Block isn't configured	The instrument has defaulted to a 96-well configuration.	Call Technical Support.
Block initialized	Block module has been re-initialized.	Call Technical Support.
Block version unknown, update firmware	The calibration data in the block is not recognized by the firmware.	Upgrade the firmware.
Bus Error	System error.	Call Technical Support.
Calibration battery RAM initialized	Calibration lost. Instrument may not perform to specification.	Call Technical Support.
Can only enter an infinity hold at end	A method can only have a HOLD segment with an infinity hold as the last segment in a method. This message occurs when you try to enter an infinity hold segment in the middle of a method.	Assign finite time segments to holds within a method.
Can't allocate timer	System error.	Call Technical Support.
Delete your methods first	User tried to delete a user name that has methods stored under it.	Delete or transfer the associated methods before deleting a user name.
Enter a name or CANCEL	You did not enter at least one character on the User Name screen before pressing the Accept key.	Enter the user name to which the desired method is assigned.
Enter oligo sequence	Incomplete TmCalc data.	Enter a value in the P1P2 fields of the TmCalc.
Enter user and method names or CANCEL	You did not enter a user name and a method name before storing a method.	Specify the method name and choose a user to store a method.
FATAL – Block shut off by hardware	Block thermal runaway.	Call Technical Support.
FATAL – Block thermal runway	Fatal error.	Turn off system. Call Technical Support.
FATAL – Cover shut off by hardware	Heated cover thermal runaway.	Call Technical Support.
FATAL – Heat sink is too hot	Ambient conditions may be too warm.	Call Technical Support.
FATAL – Heat sink sensor failure	System error.	Call Technical Support.
FATAL – Heated cover thermal runaway	Fatal error.	Turn off system. Call Technical Support.

Table 8-1 Error Messages *(continued)*

Message	Description	Recommended Action
FATAL – Sample block sensor failure	Fatal error.	Turn off system. Call Technical Support.
Fatal – Stack Overflow	A warning or error message that displays which task stack overflowed. The warning message is issued when the stack has reached within 10% of overflowing.	Call Technical Support.
Field is full	You tried to enter more data in a field than the field size allows.	Reenter data within the specifications of the field.
Heated cover sensor failure	The heated cover sensor failed.	Call Technical Support.
Infinity hold not allowed in cycle	A method can have an infinity hold segment as the last segment in the method. This message occurs when you tried to enter an infinity time in a CYCLE segment.	Use finite values for cycle segments within the method.
Invalid password/pin#	You entered an incorrect PIN#.	Enter the correct PIN#.
LCD screen timed-out	System error. Display screen and firmware have a faulty connection.	Call a Technical Support.
List of user names is full	The maximum number of users has been entered into the system.	Delete unused user names.
Maximum of 6 segments allowed	You tried to insert more than six temperature control parameters into a hold or cycle.	Do not assign more than six hold or cycle parameters to a method.
Method battery RAM initialized	Stored methods have been reset due to hardware or software failure. Not all methods may be lost.	Check method directory. Call Technical Support.
Method requires at least one segment	You deleted all temperature control parameters in a method. A method must have at least one time and temperature parameter.	Review and correct the method to include the temperature parameter(s).
No seconds in time field	You did not include seconds in the time field.	Include seconds when entering the time.
Not enough method memory left	This error occurs: ♦ When you attempt to exceed the limit of 137 methods. ♦ When you attempt to store or create a new method which is larger than the available storage space.	♦ Determine how much storage memory is available on the instrument or PC card. ♦ Delete or store rarely used methods elsewhere.
Not implemented yet	The feature is not implemented in the current firmware version.	Upgrade firmware when the new version is available from Applied Biosystems.
PC card and Flash do not verify	The PC (upgrade) card and instrument memory do not match. Firmware upgrade unsuccessful.	Call Technical Support.
PC card does not contain valid data	The PC card being used to upgrade the instrument does not contain a valid program.	Call Technical Support.

Table 8-1 Error Messages *(continued)*

Message	Description	Recommended Action
Preferences battery RAM initialized	User configuration has been reset due to software error.	Call Technical Support.
Printer not responding	The printer has been disconnected or is off line.	Check printer connections and power switch.
Remove infinity hold first	A method can have an infinity hold segment as the last segment in the method. This message occurs when a user tries to add a segment after one which contains an infinity hold.	Add segments prior to the post-PCR infinity hold.
Setpoint could not be reached	♦ The instrument could not reach a temperature parameter set by the user. The unit has a Peltier or power amplifier failure. ♦ Ambient conditions may be out of recommended range.	Call Technical Support.
SYSTEM ERROR invalid pointer	System error.	Call Technical Support.
Tm temperature out of range	Tm out of range.	Check input value and retry. Call Technical Support.
User name already defined	You entered a user name that already exists.	Do not duplicate user names.
WARNING: Block version unknown	Some data in the block is unrecognized. Instrument operation will not be effected.	Upgrade firmware.
Watchdog timeout	Software failure.	Call Technical Support.
Write to block failed	Information written to the Interchangeable Sample Block Module has failed.	Call Technical Support.
Write to default block failed	System error. Write to memory in block failed.	Call Technical Support.
Write to default Xicor failed	System error. Write to memory in the block failed.	Call Technical Support.
Write to Xicor failed	Information written to the Interchangeable Sample Block Module has failed.	Call Technical Support.
Valid range is	You entered a number out of range. The message include the valid range limits.	Reenter a value within the parameters of the field.

Troubleshooting Information

Troubleshooting Table Refer to the following table for a description of potential problems, possible causes, and recommended actions that you should take.

Table 8-2 Troubleshooting Information

Problem	Possible Causes	Check and/or Remedy
Control panel not responding	Keypad failure.	Run keypad diagnostic. Call Technical Support.
Cooling rate too slow	♦ Ambient temperature is too warm. ♦ Peltier failure.	♦ Move instrument to well-ventilated location(15-30°C). ♦ Run rate test diagnostic. Call Technical Support.
Cycling time too long	Peltier failure.	Run cycle test diagnostic
Displayed temperature does not match specified temperature	Instrument may require calibration.	Run the Temperature Verification test.
Heated cover not responsive	Heated cover failure.	Call Technical Support.
Heating rate too slow	♦ Peltier failure.	Run Rate Test diagnostic. Call Applied Biosystems Technical Support.
Instrument can't reach high or low temperature range	♦ Ambient temperature is too warm. ♦ Peltier failure.	♦ Run Rate Test diagnostic. ♦ Run Cycle Test diagnostic. Call Technical Support.
Instrument making too much noise	Fan failure.	Check for sidevent obstructions.
No beep	♦ Run time beeper disabled. ♦ Beeper failure.	Check Run-Time Beep configuration. Call Technical Support.
No screen display No response when you turn the instrument on	♦ Fuse blown. ♦ Not connected to power source. ♦ Interchangeable module not installed correctly.	♦ Is power switch ON? ♦ Is power cord connected? ♦ Check fuses.
Printer fails	♦ Incorrect printer configuration. ♦ Incorrect printer cable.	♦ Check printer settings: baud rate = 9600, no parity, one stop bit, eight data bits. ♦ Purchase Applied Biosystems printer cable.
Instrument cooling fan does not make whirring sound	♦ Fuse blown. ♦ Not connected to power source. ♦ Interchangeable module not installed correctly.	♦ Is power switch ON? ♦ Is power cord connected? ♦ Check fuses.

Instrument Specifications

A

Overview

About This Appendix

This appendix describes the dimensions, power, and electrical specifications for the GeneAmp® PCR System 9700 system, including the control panel, sample temperature information, and printer specifications.

In This Appendix

The following topics are covered in this appendix:

Topic	See page
System Specifications	A-2
Control Panel Specifications	A-4
Sample Temperature Information	A-4
Printer Specifications	A-4

System Specifications

Dimensions

The following tables list the footprint and the weight of the instrument with the 96–Well or 60–Well sample block module.

Footprint (With Sample Block Module Installed)

Height	26 cm (10 in)
Width	28 cm (11 in)
Depth	41 cm (16 in)

Note You must provide sufficient space around the instrument for unrestricted air circulation.

Weight

Base Module	8.6 kg (19 lbs)
96-Well Gold/Aluminum Sample Block Modules	3.2 kg (7 lbs)

Note See the sample block module user's manual for physical information on a module.

Power Configurations

There is one version of the instrument. The power requirements of the instrument under various power configurations are:

VAC ~100/120	8 AMP T (5x20 mm) or 8 AMP Slow Blow (3 AB)	50/60 Hz Use 250 V fuses
VAC ~220/230/240	8 AMP T (5x20 mm)	Max Power 725 VA

Electrical Requirements

IMPORTANT You must be able to disconnect the main power supply to the instrument immediately if necessary.

In areas where the supplied power is subject to voltage fluctuations exceeding +/-10% of the nominal value, a power line regulator may be required. High or low voltages can have adverse effects on the electronic components of the instrument. The following table specifies the electrical operating range for the instrument in various parts of the world. Select appropriate fuse configuration based on the voltage used.

WARNING In Japan, the unit must have a dedicated 220-volt outlet! The unit will not operate properly with a 100-volt outlet.

Location	Voltage (VAC)[a]	Frequency	Amperage (A) Nominal
Japan	220 ±10%	50/60 Hz ±1%	3.16
USA/Canada	120 ±10%	50/60 Hz ±1%	4.20
Europe (pre-1992)	220 ±10%	50/60 Hz ±1%	3.16
EC	230 ±10%	50/60 Hz ±1%	3.14
UK (pre-1992)	240 +6%/–10%	50/60 Hz ±1%	3.12
Australia	240 +6%/–10%	50/60 Hz ±1%	3.12

a. Acceptable AC line voltage tolerances: 100, 120, 220, 230 ±10%; 240 VAC +6%/–10%, 50/60 Hz ± 1%.

Note The Volt-Amp number for this instrument is 725Volt Amps.

Control Panel Specifications

Display Screen The display screen is a 7 x 40 character display with a graphics mode of 60 x 240 pixel resolution.

Keys The instrument control panel consists of a display screen and 22 keys. The keys are:

- Function keys
- Arrow keys
- Stop key
- Enter key
- 10 number keypad

Sample Temperature Information

Temperature The following table lists sample temperature information.

Note Sample temperatures are displayed in degrees Celsius to the nearest 0.1 °C.

Sample Temperature Range	4.0 to 99.9 °C.
Temperature Calibration	Traceable to National Institute of Standards and Technology (NIST).

Printer Specifications

Serial Interface Board Specifications The instrument can use any printer with a serial (RS-232C) interface board with the following parameters.

Baud Rate	9600
Parity	NONE
Data Bits	8
Stop Bits	1

Cable Part Number Connect the printer to the Instrument port with printer cable part number N805-1326.

Supplied Methods

B

Overview

About this Appendix

This appendix provides information about the methods that are supplied with the GeneAmp® PCR System 9700.

In This Appendix

The following topics are covered in this appendix:

Topic	See page
About the Methods	B-2
AmpliCycle Sequencing	B-2
AmpliTaq Gold DNA Polymerase	B-2
BigDye Terminators	B-3
General PCR	B-3
LMS2	B-4
Time Release PCR	B-4
Touchdown PCR	B-5
XL PCR	B-5

About the Methods

Introduction

The instrument supplies you with eight pre-coded methods stored under the user name <<pe>>.

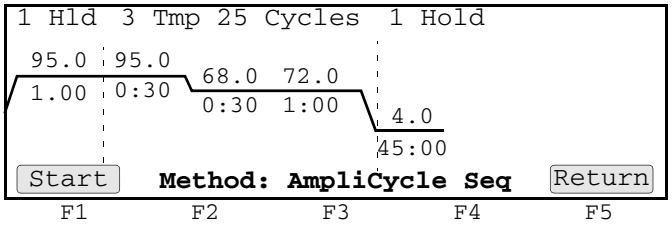
The eight pre-coded methods are:

- AmpliCycle® Sequencing
- AmpliTaq Gold® DNA Polymerase
- BigDye® Terminators
- General PCR
- LSM2
- Time Release PCR
- Touchdown PCR
- XL PCR

See the following sections for detailed information about the methods.

AmpliCycle Sequencing

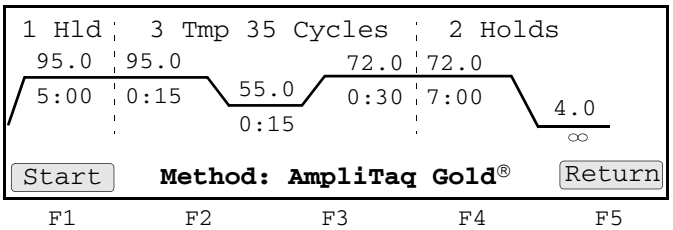
Cycle sequencing has revolutionized Sanger sequencing of PCR products and other DNA templates. The AmpliCycle Sequencing protocol is for 3-temperature cycling and for achieving clean sequence ladders from femtomole amounts of template.



This cycle sequencing process and the benefits of AmpliTaq® DNA Polymerase, CS, are described in the product insert for the AmpliCycle® Sequencing Kit (P/N N808-0175).

AmpliTaq Gold DNA Polymerase

The AmpliTaq Gold protocol specifies a 5-minute pre-PCR heat step, required for the activation of AmpliTaq Gold® DNA Polymerase. This additional step provides seamless “hot start” PCR and replaces labor intensive methods such as manual hot start or wax bead-mediated hot start techniques.

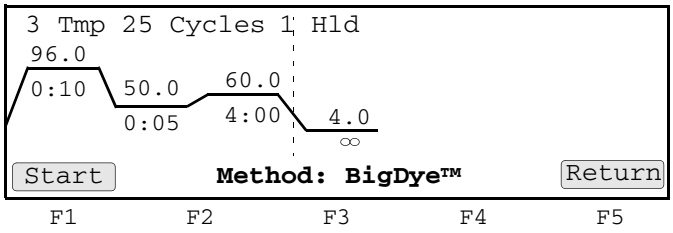


Utilizing hot start techniques helps to minimize the formation of primer-dimers or non-specific products, thus increasing specificity and sensitivity of PCR.

You can find further information on AmpliTaq Gold DNA Polymerase in the product insert (P/N N808-0241) or at the Applied Biosystems website.

BigDye Terminators

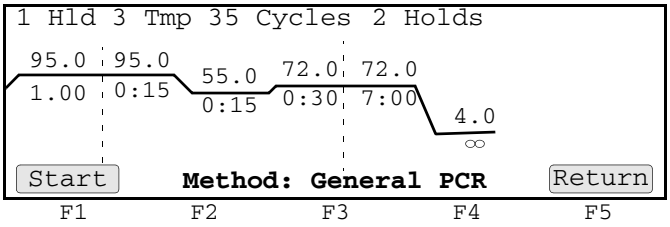
The BigDye® method consists of cycle sequencing parameters for dideoxy (Sanger) terminator sequencing using ABI PRISM® BigDye® Terminator Cycle Sequencing Ready Reaction Kits (available from Applied Biosystems). It consists of 25-cycle, three-temperature cycle sequencing followed by an infinite hold at 4 °C.



The pre-coded conditions in the BigDye® method are optimized for AmpliTaq® DNA Polymerase, FS and the cycle sequencing reagents supplied with the BigDye terminator kits. This process is further described in the *ABI PRISM BigDye Terminator Cycle Sequencing Ready Reaction Kits Protocol* (P/N 4303237).

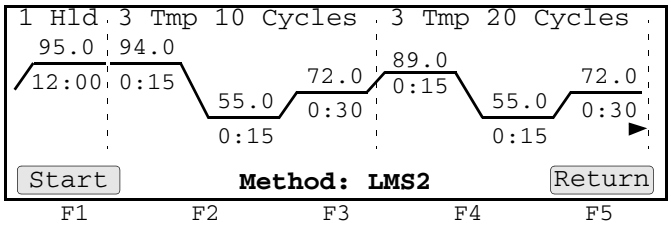
General PCR

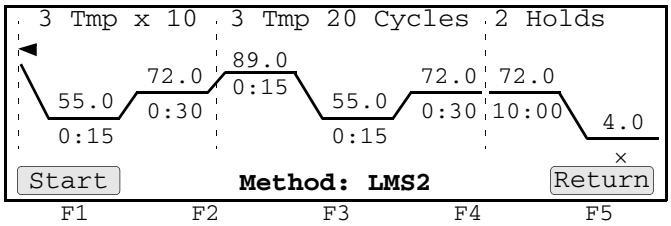
The General PCR method is a basic one and can be easily modified with both pre- and post-PCR holds.



LMS2

The GeneAmp PCR System 9700 software includes a pre-coded LMS2 method for ABI PRISM® Linkage Mapping Set Version 2 (LMS2) thermal cycling. Linkage Mapping Set 2 employs over 400 fluorescent-labeled PCR primer pairs for analysis of select microsatalite loci from the Généthon human linkage map.[1,2,3] The following figures illustrate the thermal cycling profile for the LMS2 method.



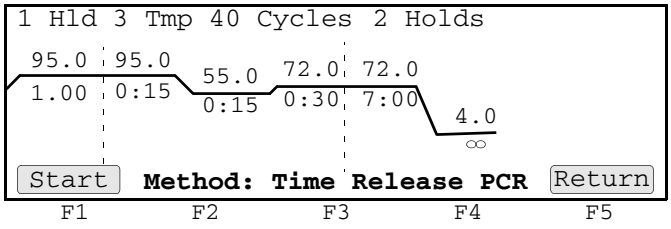


This supplied method consists of an initial hold at 95.0 °C, two sets of three temperature cycles, followed by two additional holds at 72.0 °C and 4.0 °C.

See the *ABI PRISM Linkage Mapping Set Version 2 User's Manual* (P/N 904999) for comprehensive information on PCR amplification conditions, electrophoresis conditions, detection, and data analysis.

Time Release PCR

The Time Release PCR method is designed for use with AmpliTaq Gold DNA Polymerase. The enzyme is activated more slowly than with the AmpliTaq Gold method. Here the pre-PCR hold is only 1 minute, and the number of cycles is increased to 40.



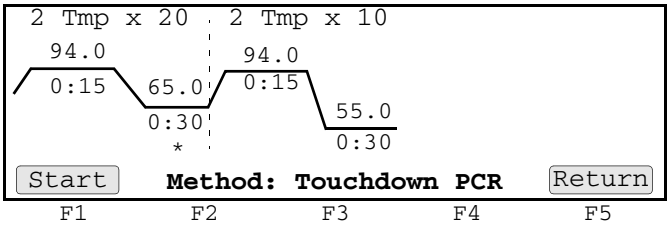
1. Weissenbach, J. *et al.* 1992. A second-generation linkage map of the human genome. *Nature* 359:794–801.

2. Gyapay, G., *et al.* 1994. Généthon Human Genetic Linkage Map. *Nature Genet.* 7:246–339.

3. Dib, C., *et al.* 1996. *Nature* 380: 152–154.

Touchdown PCR

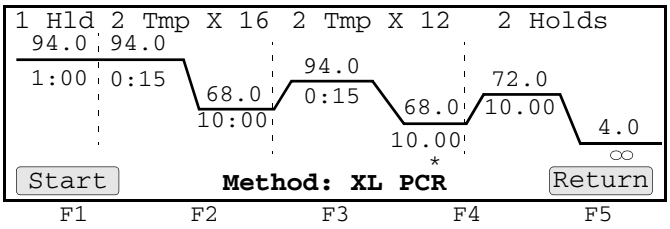
When the optimal annealing temperature is unknown, one strategy, touchdown PCR, incrementally decreases the annealing temperature in early cycles in order to maximize the yield of specific products.



This supplied method has an initial annealing temperature (65 °C) that incrementally decreases by an additional 0.5 °C in each of the first 20 cycles, followed by 10 cycles at 55 °C.

XL PCR

XL PCR is the protocol specified for amplification of 5 kb-40 kb PCR products, using r*Tth* DNA Polymerase, XL, and unique reaction conditions.



This protocol uses two-temperature cycling (94 °C for 15 seconds; 68 °C for 10 minutes) and invokes a 15 second AutoX (automatic segment extension) for the anneal/extend step in the last 12 cycles.

By providing longer templates, XL PCR complements technologies for rapid, long-range PCR. More complete genes can be amplified in one reaction from known expressed sequences, thus more introns can be crossed. You can use XL PCR for the amplification of the control target, a 20.8 kb product from Lambda DNA, supplied in the kit.

This process is further described in the product insert for the GeneAmp® XL PCR Kit (P/N N808-0192).

Contacting Services and Support

C

How to Obtain Services and Support

To contact Applied Biosystems Technical Support from North America by telephone, call **1.800.899.5858**.

For the latest services and support information for all locations, go to **http://www.appliedbiosystems.com**, then click the link for **Services and Support**.

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- Submit a question directly to Technical Support
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- Obtain information about customer training
- Download software updates and patches

In addition, the Services and Support page provides access to worldwide telephone and fax numbers to contact Applied Biosystems Technical Support and Sales facilities.

Limited Warranty Statement

D

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Limited Warranty

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